

fMRI Course, Day 9
Pitfalls of fMRI Analysis

Tuesday, August 9th, 2022

Before We Begin

Questions about either 1st- or 2nd-level analysis?

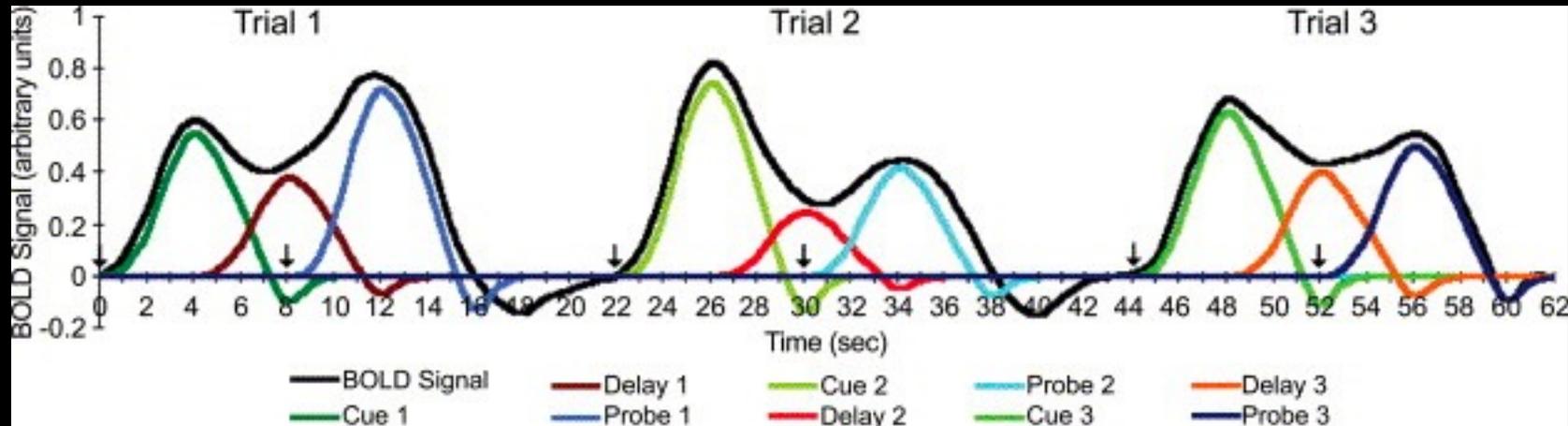
Questions about anything else?

Extra Slides from Previous Lectures

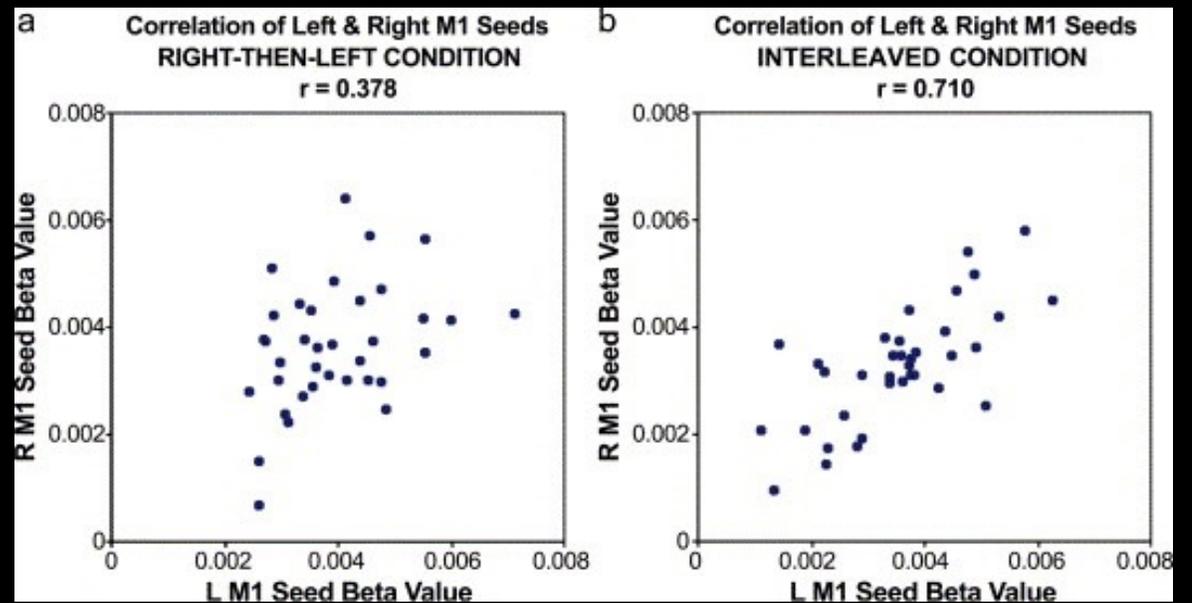
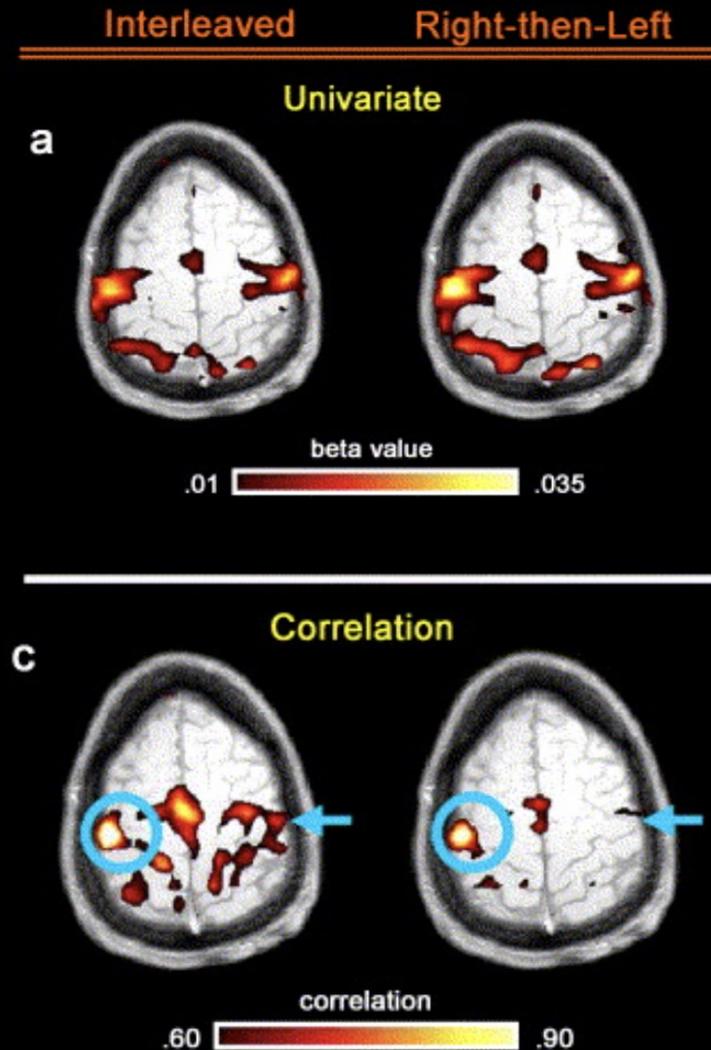
Applying the GLM to fMRI Data

Other options: Individual Modulation

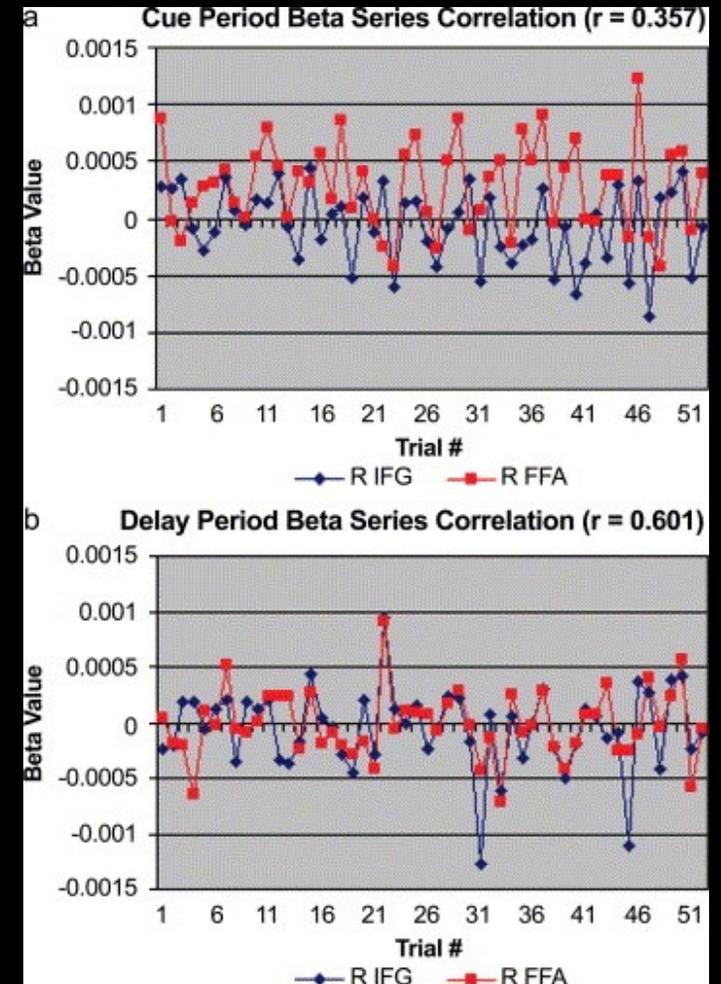
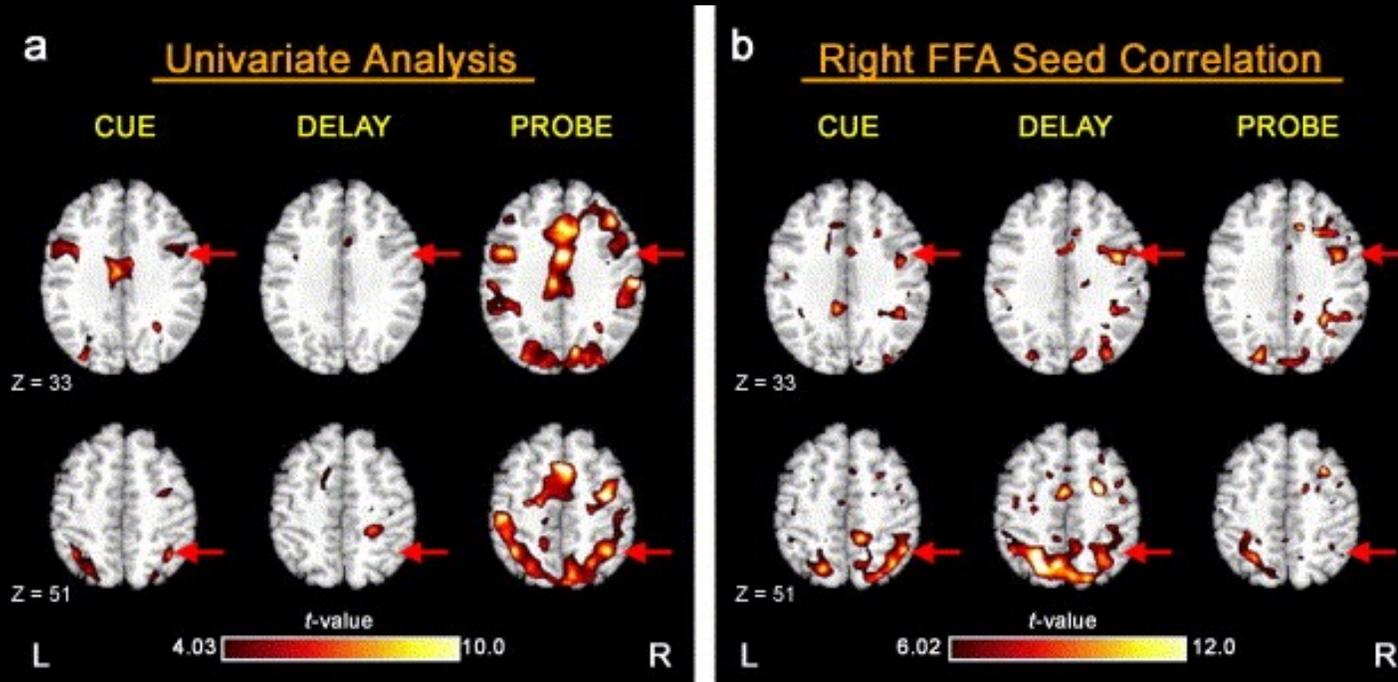
Useful for beta-series correlation, MVPA classification



Applying the GLM to fMRI Data



Applying the GLM to fMRI Data



Applying the GLM to fMRI Data

Drawback: Very tedious to implement without scripting!

For AFNI users: Can use 'IM' basis function in 3dDeconvolve

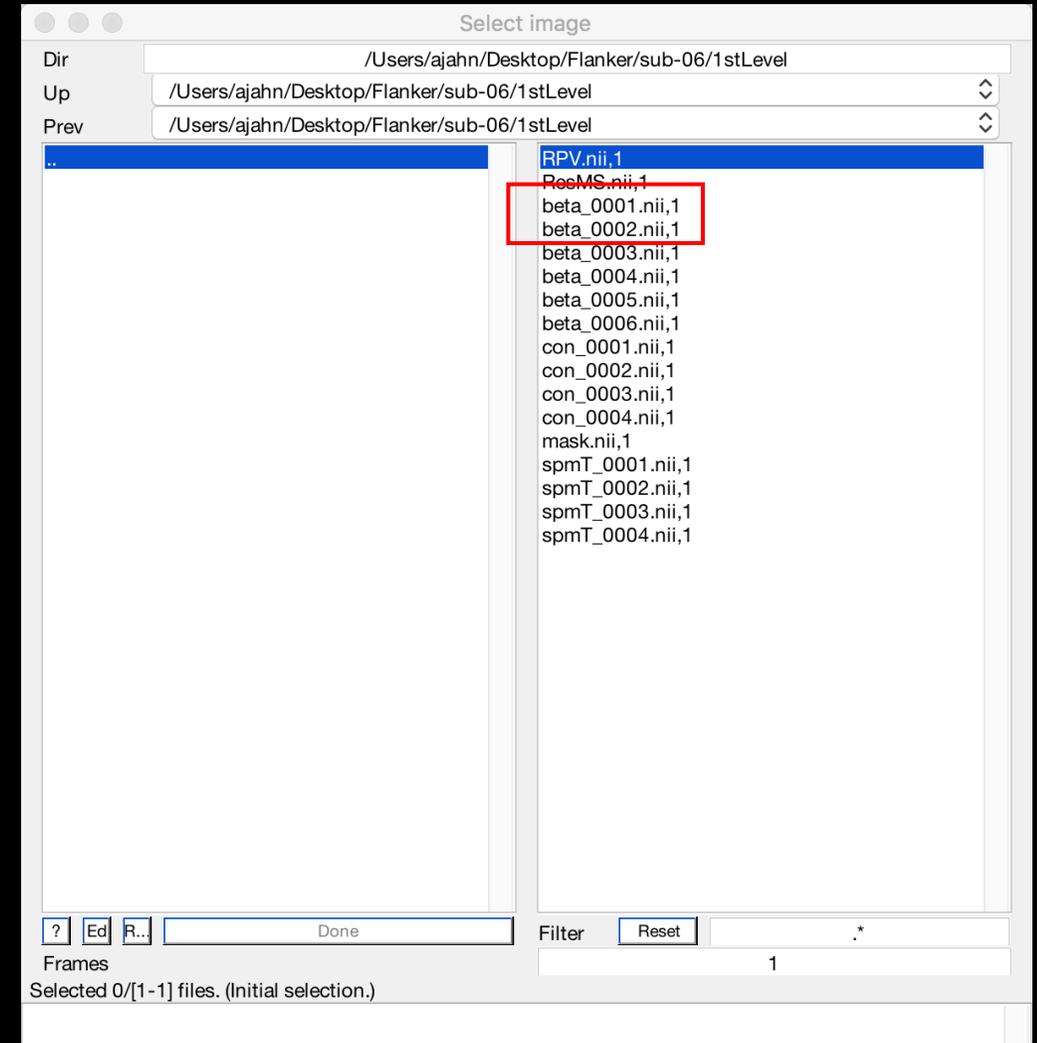
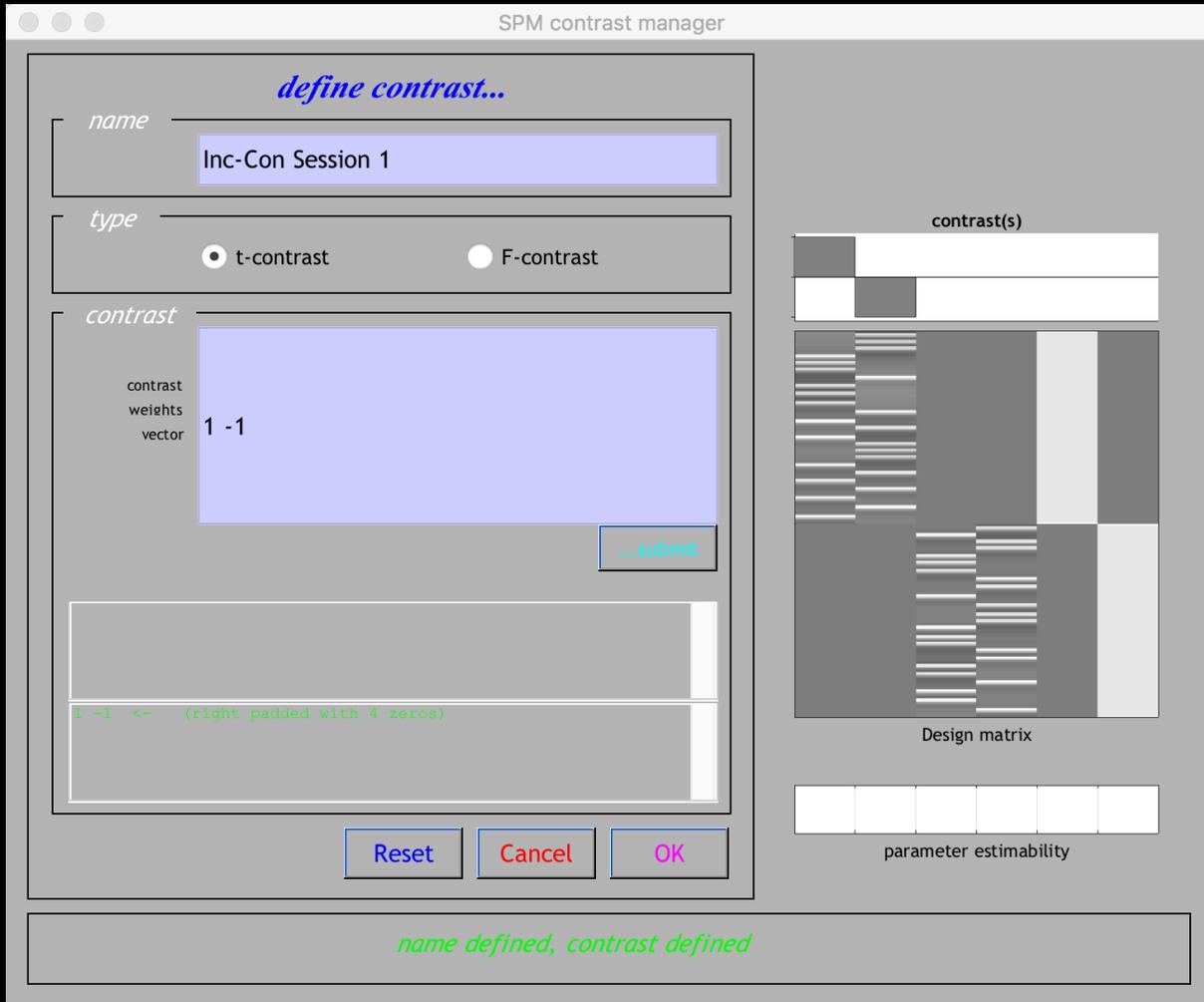
ImCalc: The Image Calculator

One of the most versatile tools is the image calculator

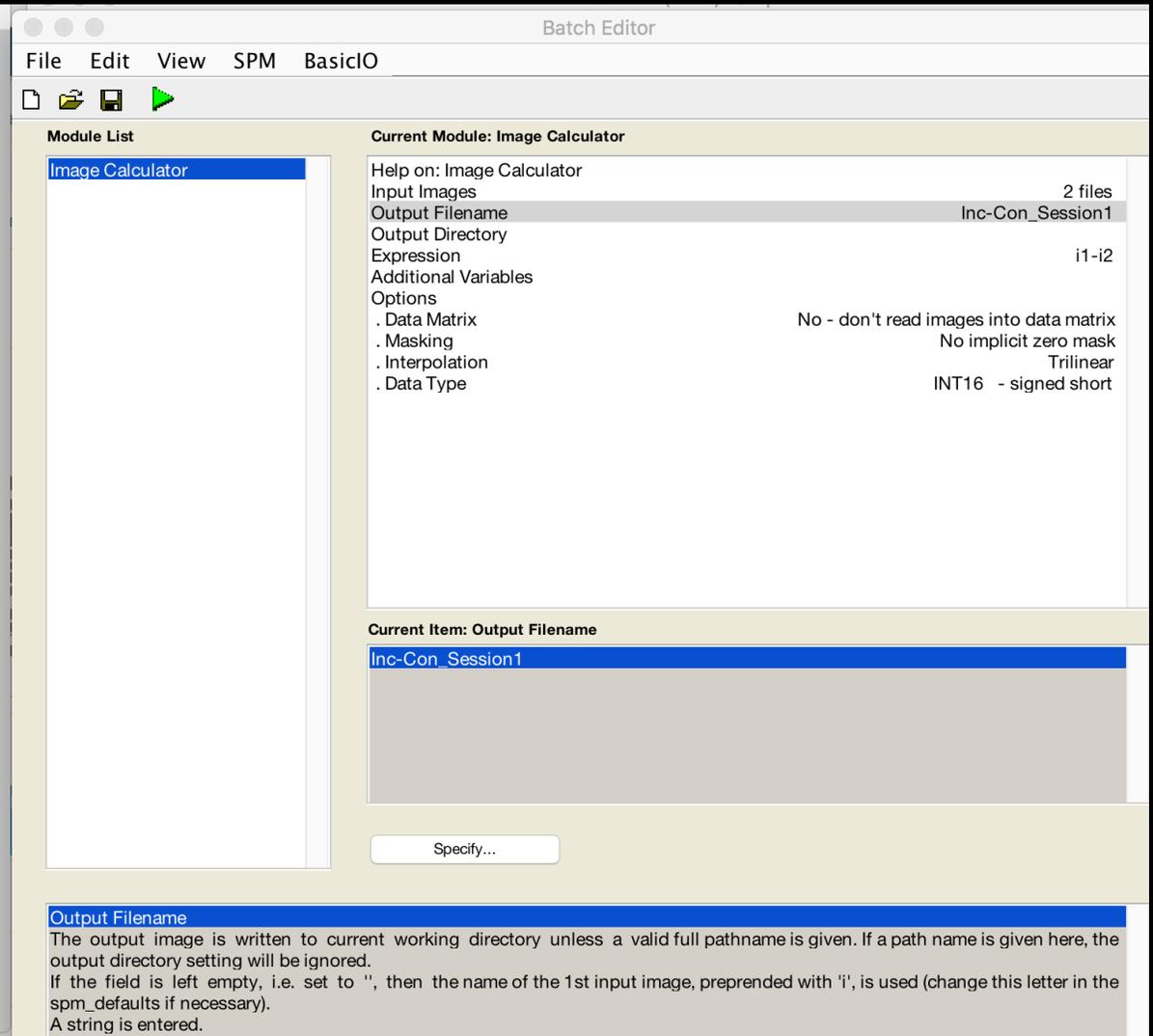
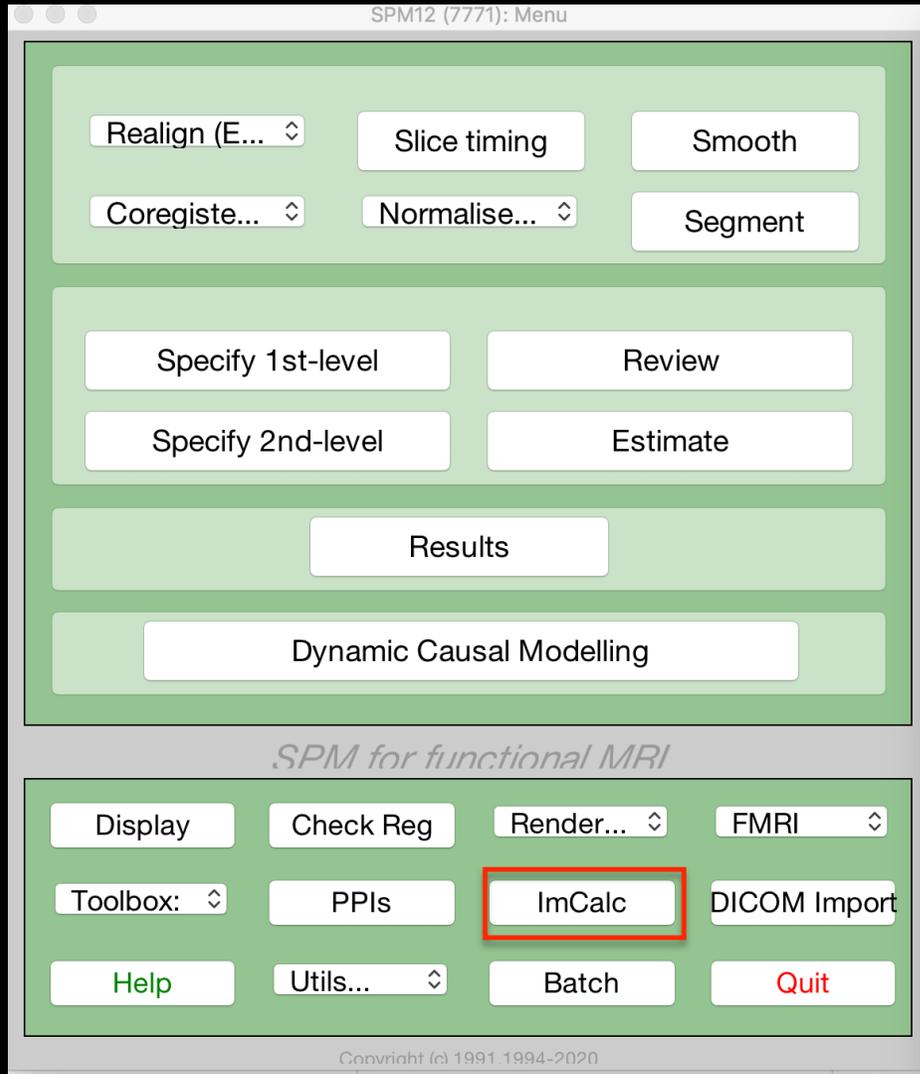
Every package has one (AFNI, FSL, MRtrix, etc.)

Simple to do basic arithmetic on a 3D image

ImCalc: The Image Calculator



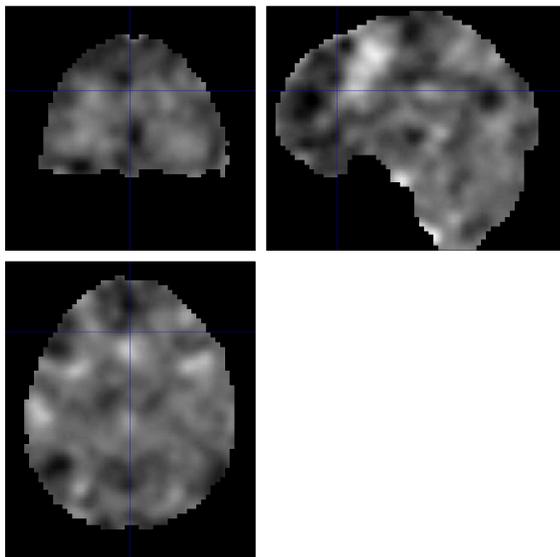
ImCalc: The Image Calculator



ImCalc: The Image Calculator

SPM12 (7771): Graphics

File Edit View Insert Tools Desktop Window SPM Figure Help



Crosshair Position Origin

mm: 0.0 30.0 30.0

vx: 27.0 48.3 34.3

Intensity: 0.000696186

right {mm}	0
forward {mm}	0
up {mm}	0
pitch {rad}	0
roll {rad}	0
yaw {rad}	0
resize {x}	1
resize {y}	1
resize {z}	1

Set Origin Reorient...

File: **./1stLevel/beta_0001.nii**

Dimensions: 53 x 63 x 52

Datatype: float32

Intensity: Y = 1 X

spm_spm:beta (0001) - Sn(1) Inc*bf(1)

Vox size: -3 x 3 x 3

Origin: 27 38.3 24.3

Dir Cos: 1.000 0.000 0.000
0.000 1.000 0.000
0.000 0.000 1.000

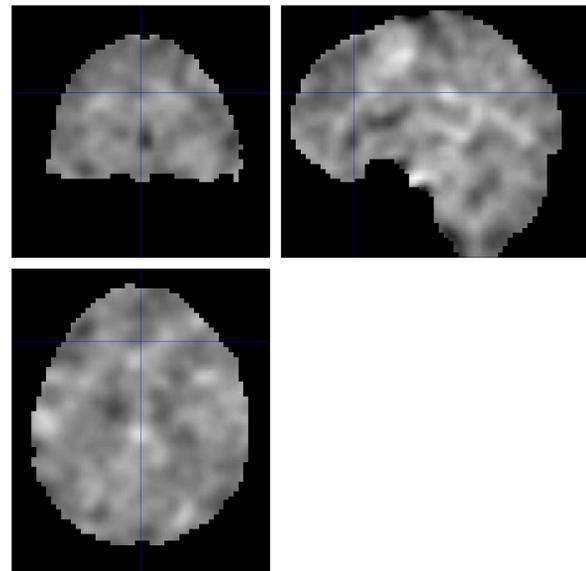
Full Volume Hide Crosshair

World Space Trilinear interp.

Auto Window Add Overlay...

SPM12 (7771): Graphics

File Edit View Insert Tools Desktop Window SPM Figure Help



Crosshair Position Origin

mm: 0.0 30.0 30.0

vx: 27.0 48.3 34.3

Intensity: -0.37464

right {mm}	0
forward {mm}	0
up {mm}	0
pitch {rad}	0
roll {rad}	0
yaw {rad}	0
resize {x}	1
resize {y}	1
resize {z}	1

Set Origin Reorient...

File: **./1stLevel/beta_0002.nii**

Dimensions: 53 x 63 x 52

Datatype: float32

Intensity: Y = 1 X

spm_spm:beta (0002) - Sn(1) Con*bf(1)

Vox size: -3 x 3 x 3

Origin: 27 38.3 24.3

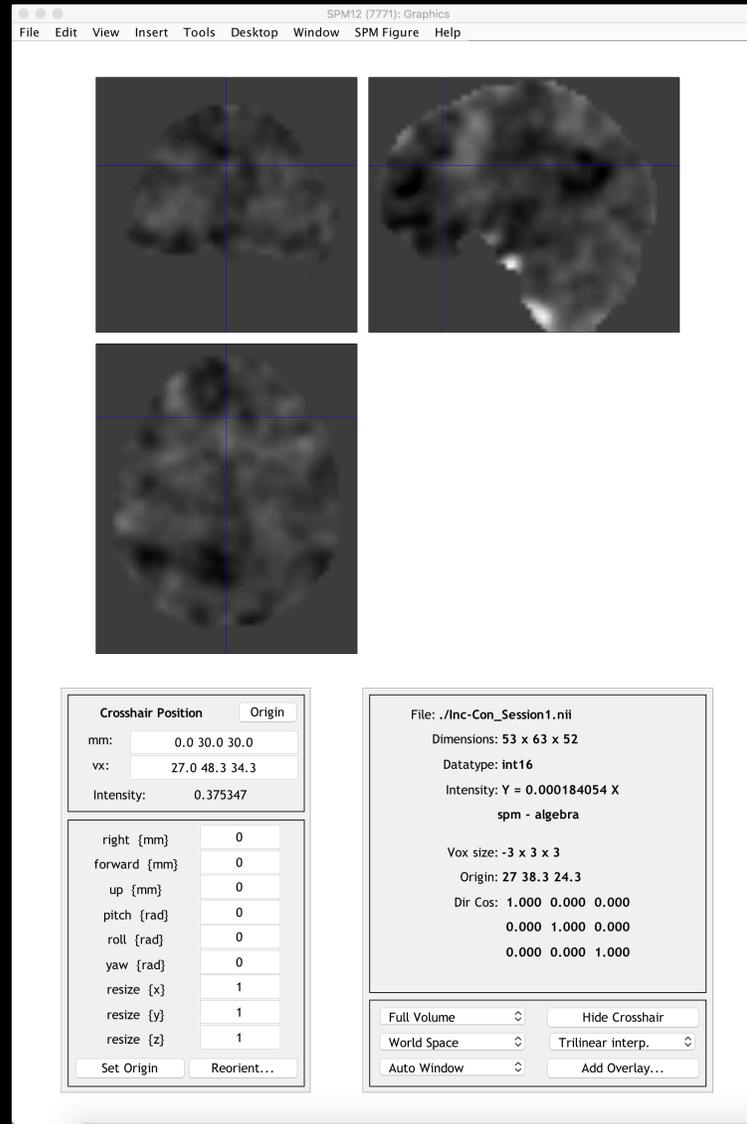
Dir Cos: 1.000 0.000 0.000
0.000 1.000 0.000
0.000 0.000 1.000

Full Volume Hide Crosshair

World Space Trilinear interp.

Auto Window Add Overlay...

ImCalc: The Image Calculator



Outline

Today: Pitfalls of fMRI Analysis

Specifically, non-independence and cluster failure

Ways to avoid these pitfalls

Articles

Scientific Standards in Epidemiologic Studies of the Menace of Daily Life

ALVAN R. FEINSTEIN

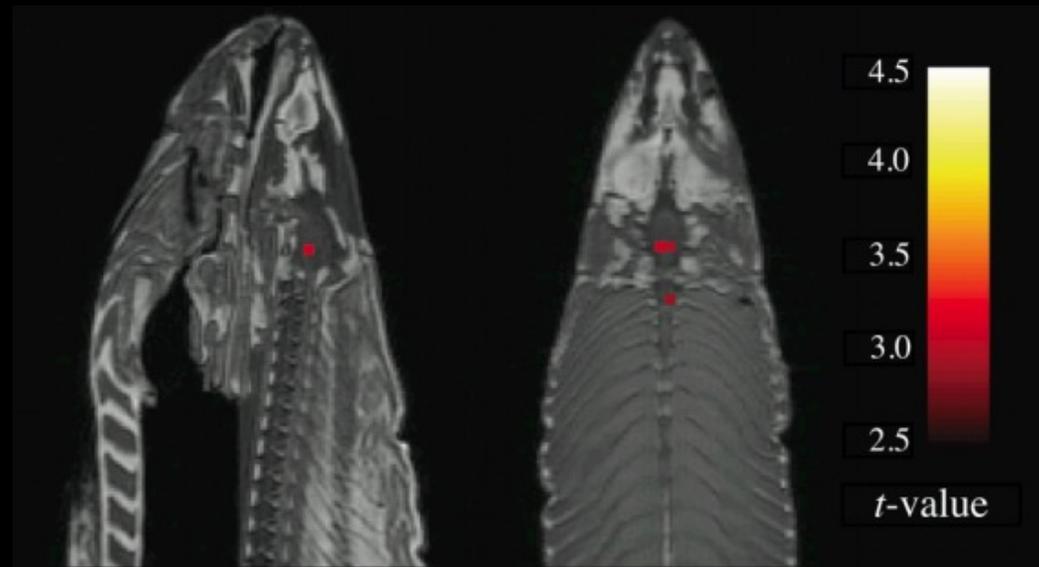
Focused on methods of epidemiological studies

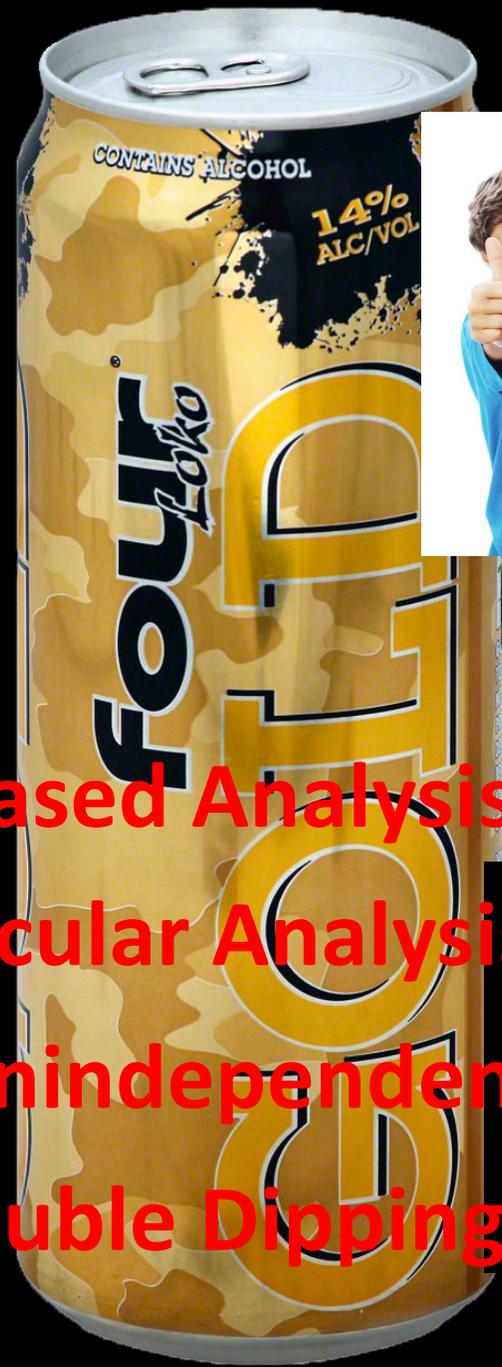
Hypothesizing after the fact

Uncorrected for multiple comparisons

Back in 2009...

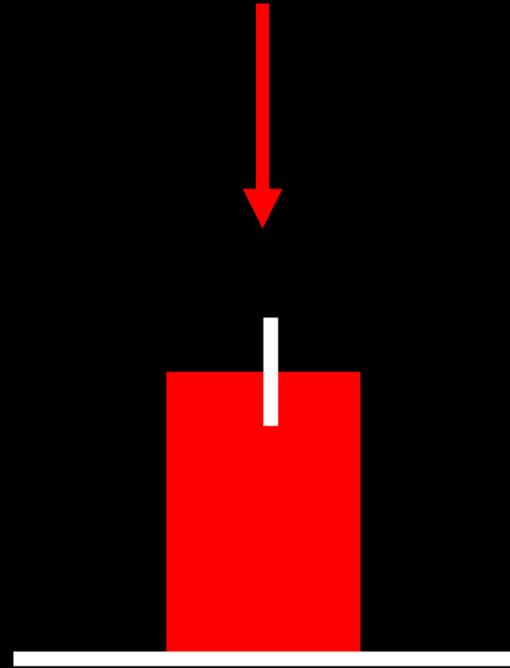
**Bennet et al.: Illustration of the
Multiple Comparisons Problem**



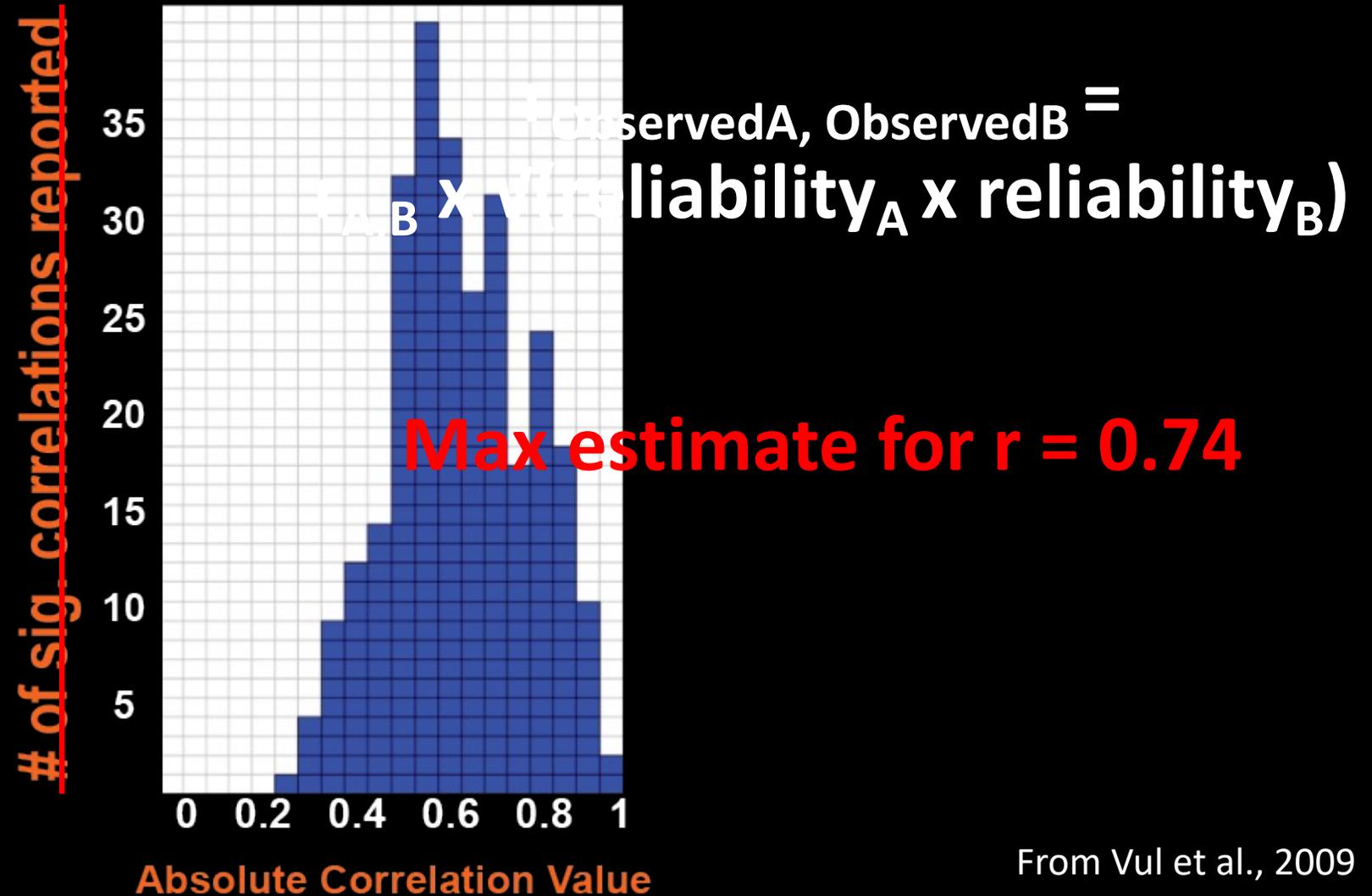


Biased Analysis
Circular Analysis
Nonindependent
Double Dipping

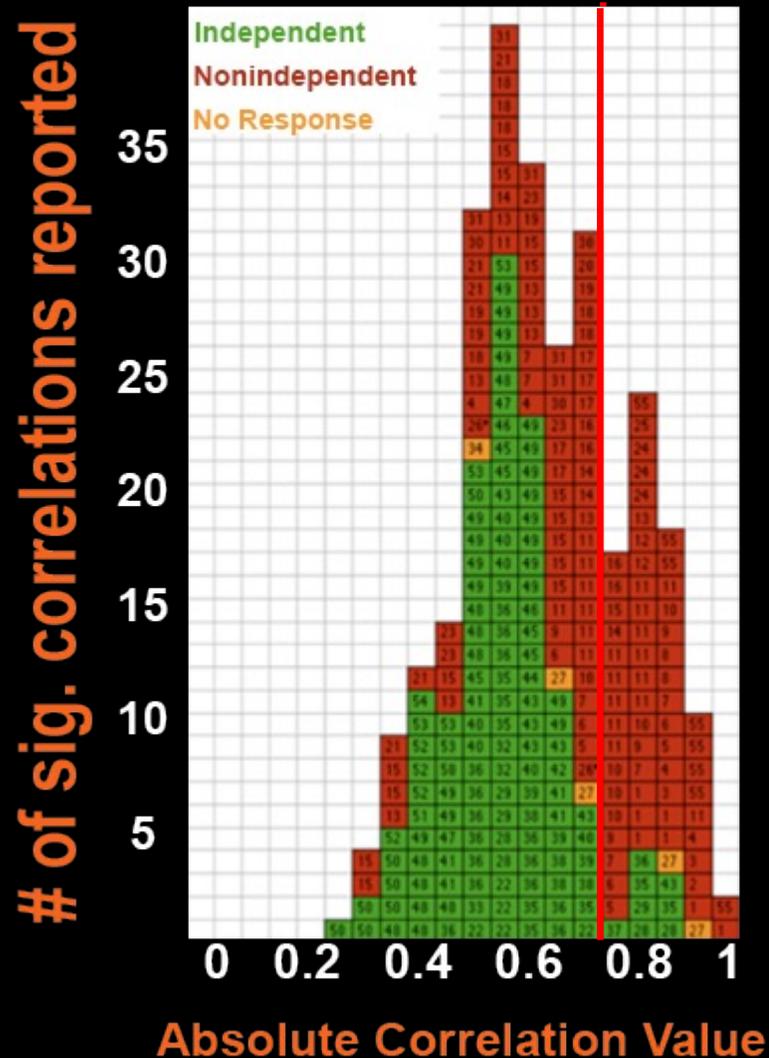
Test Score



Back in 2009...



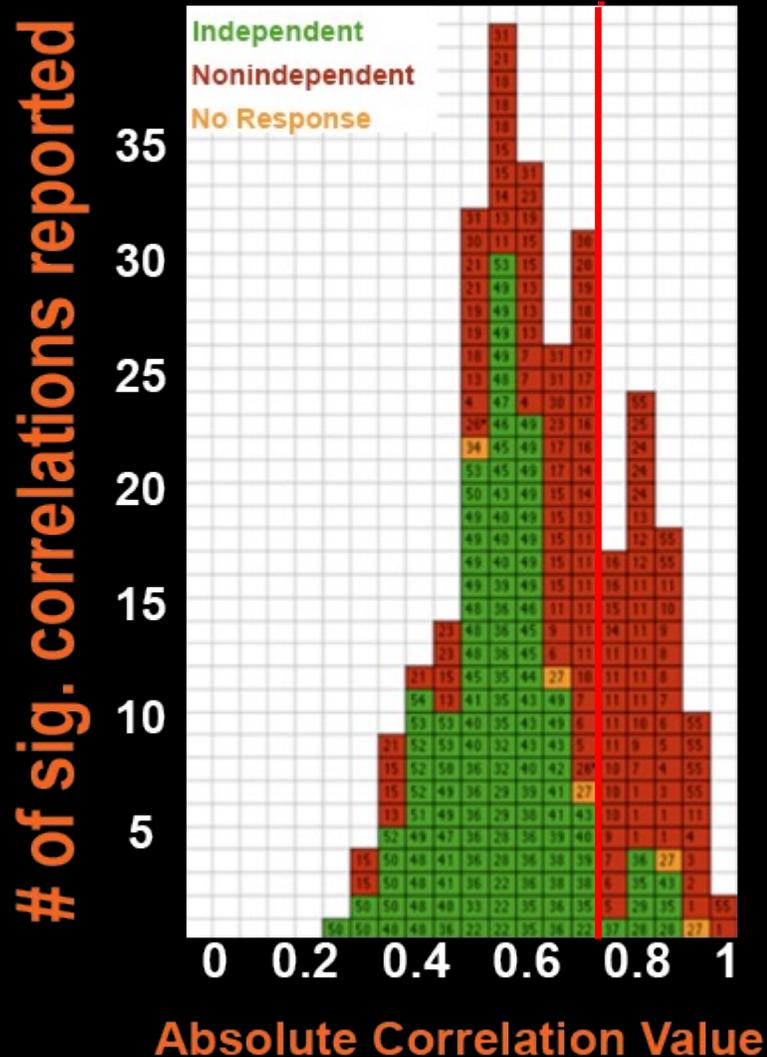
Back in 2009...



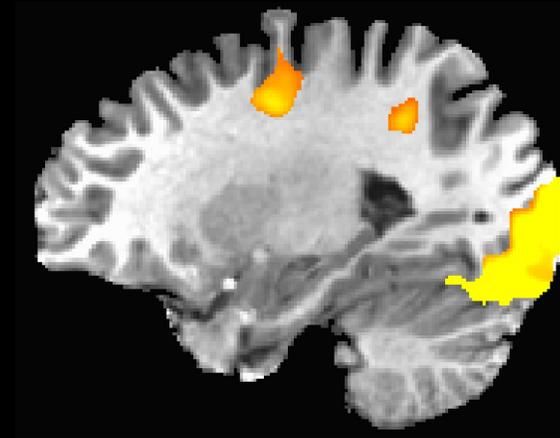
$$r_{\text{ObservedA, ObservedB}} = r_{A,B} \times \sqrt{(\text{reliability}_A \times \text{reliability}_B)}$$

Max estimate for $r = 0.74$

Vul's Interpretation



Significant voxels more likely to benefit from noise



Leads to inflated effect sizes

Objections

What if I just want to see what's going on?

My recommendation: Look, but don't publish

If included, explain how it was done,
and do *not* include error bars

Objections

But doesn't the correlation (or effect) actually exist?

Assuming multiple comparisons correction, yes

*But, magnitude is also important,
and is misleading with biased analyses*

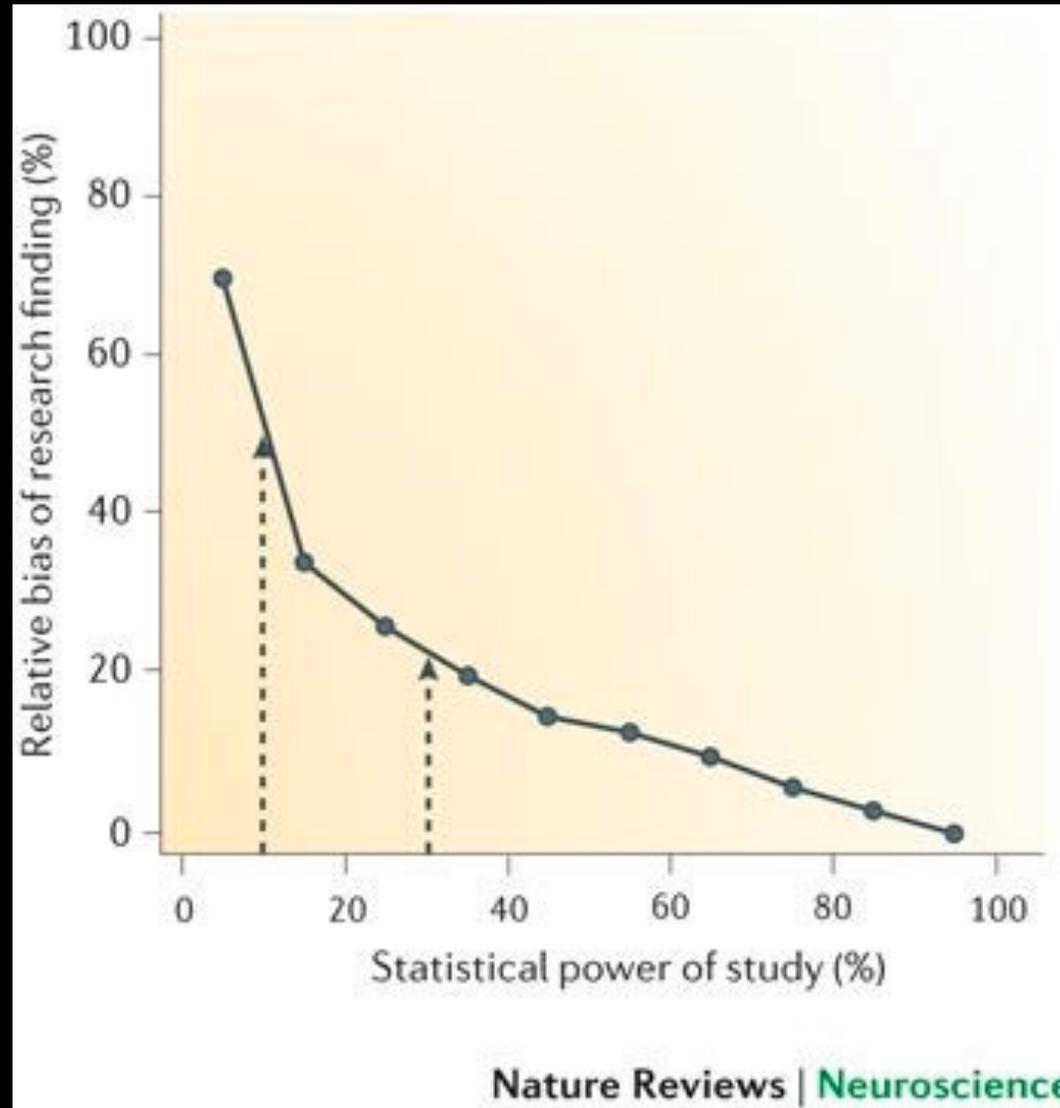
Problems with Sample Size

Only high correlations are reported

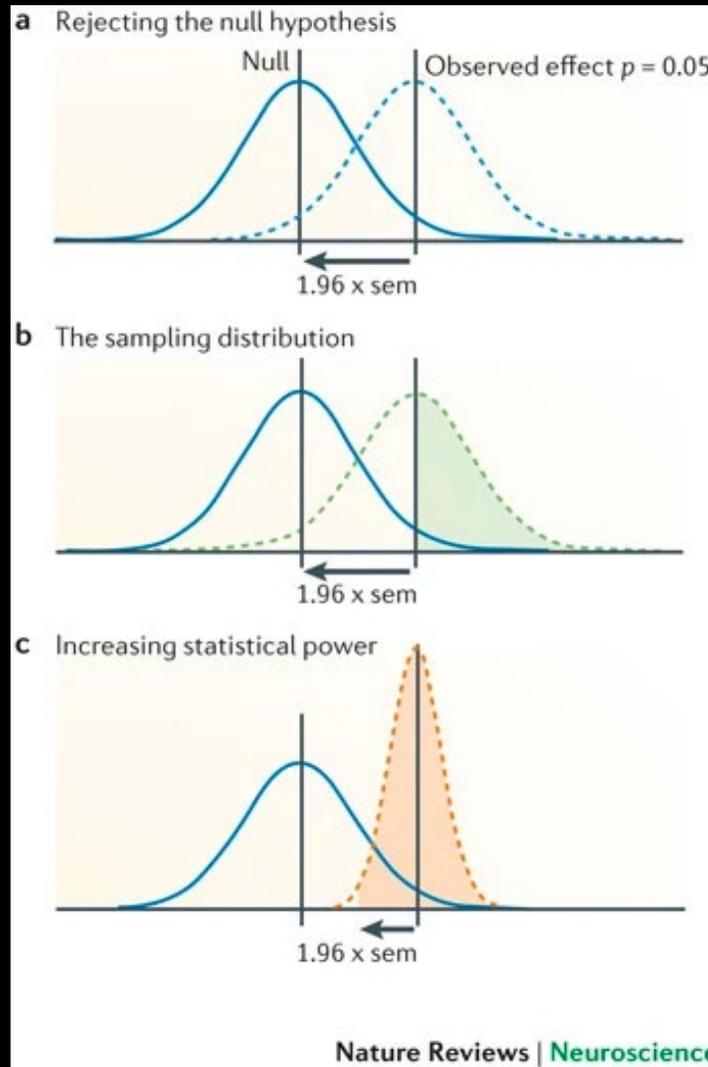
Power drops, effect size inflates

“Winner’s Curse”

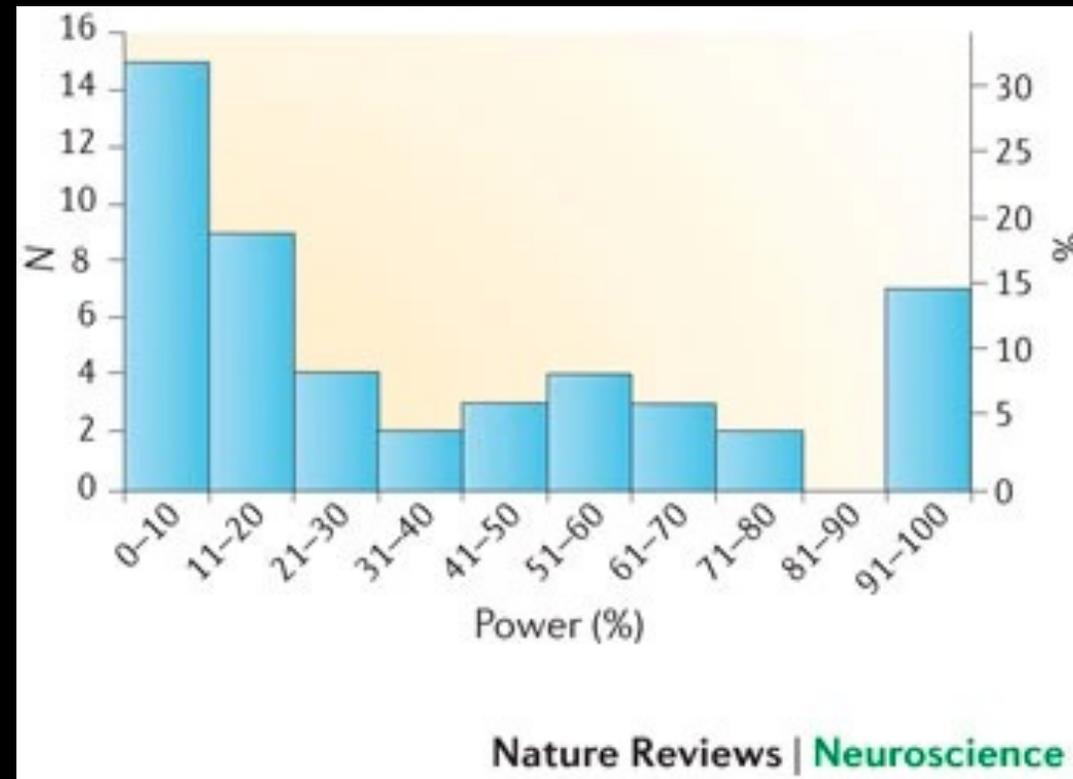
Problems with Sample Size



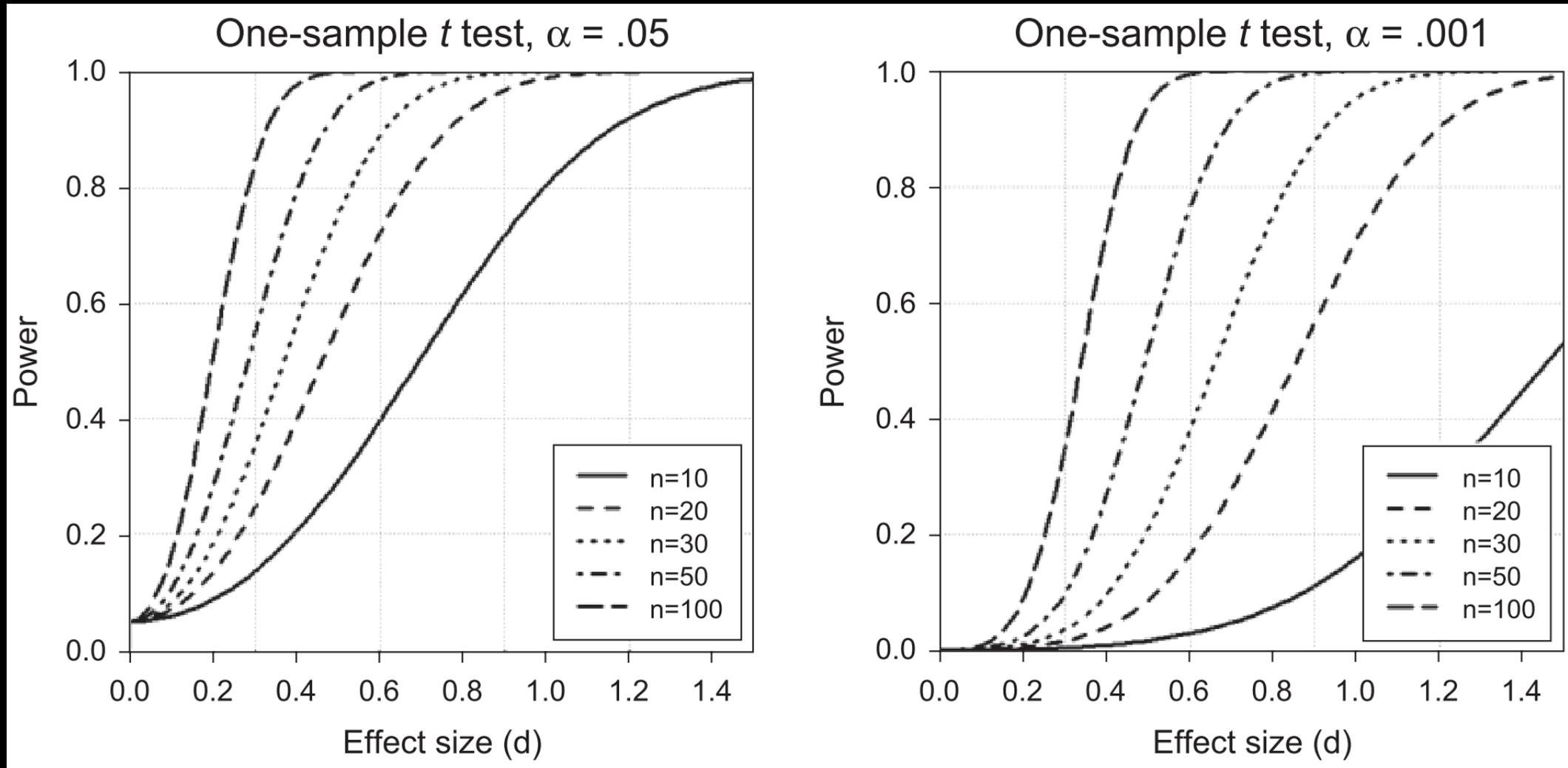
Problems with Sample Size



Problems with Sample Size



Problems with Sample Size

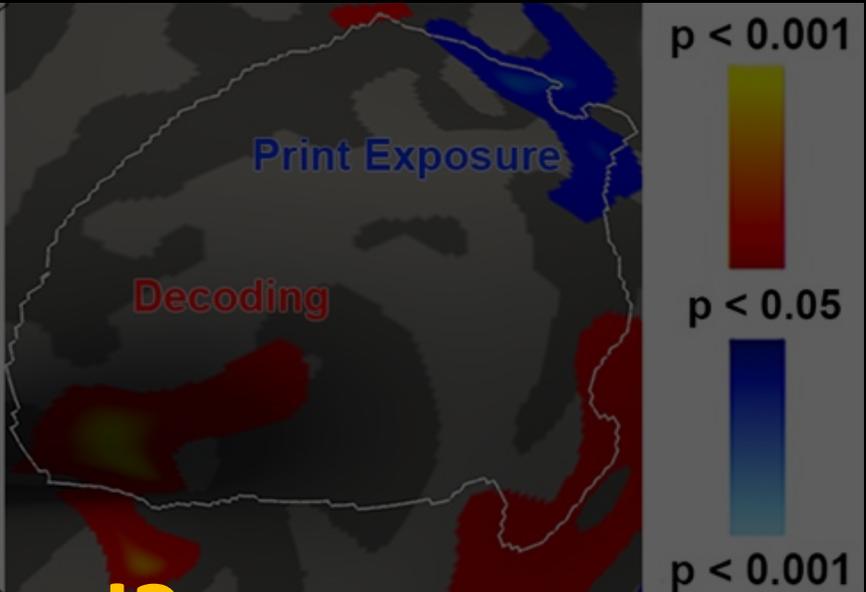


What to do?

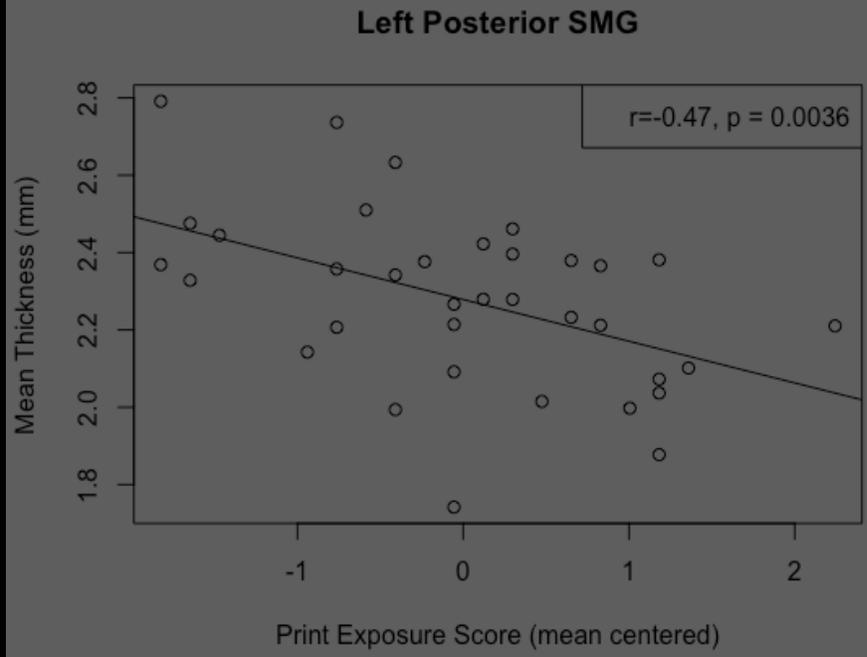
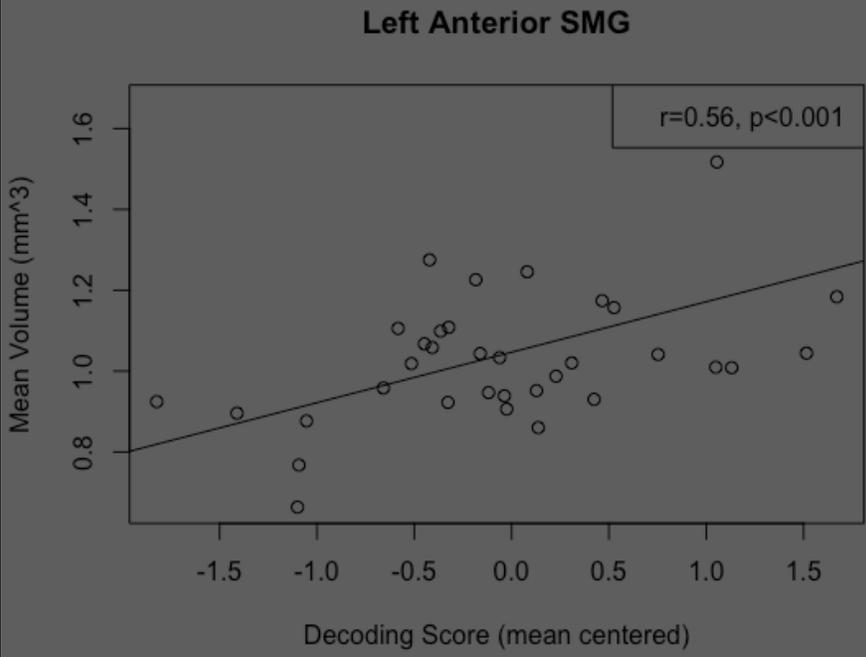
Increase N

Power analyses to estimate effect size

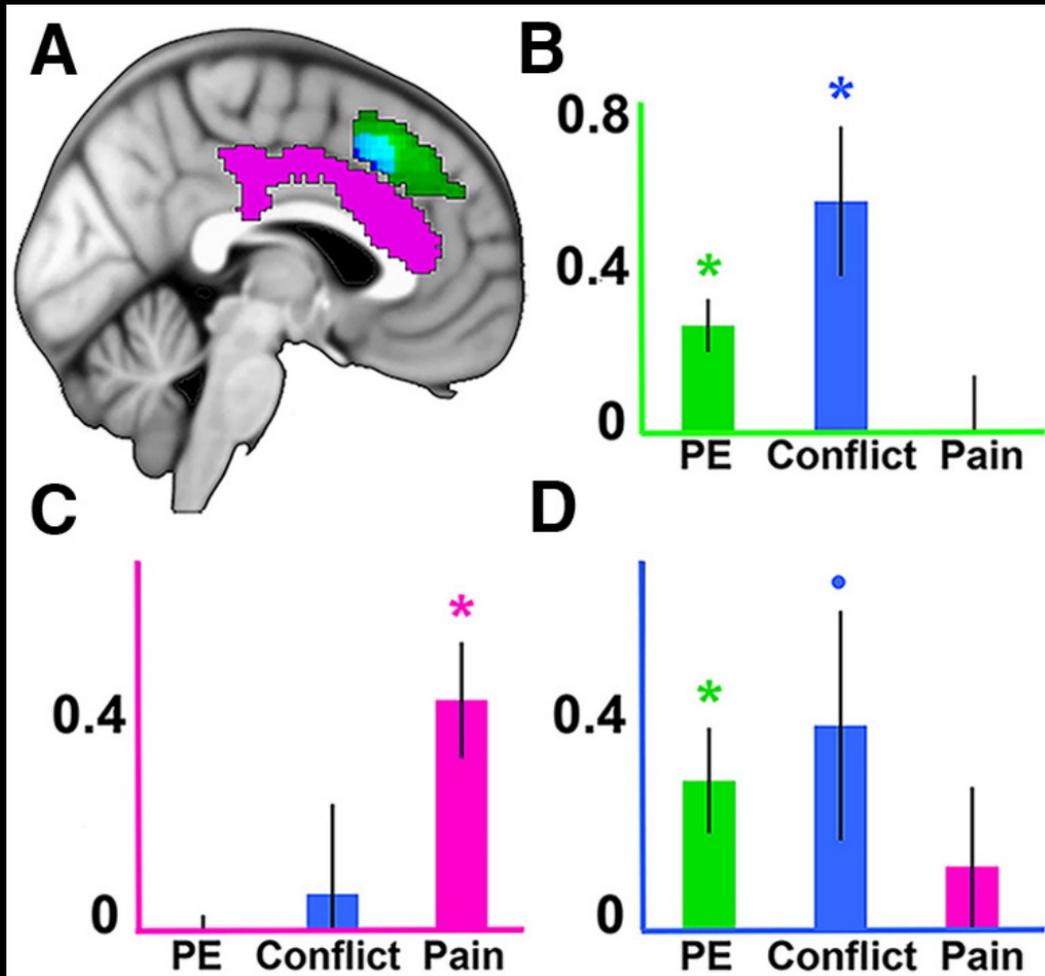
Pre-registered reports



Biased?

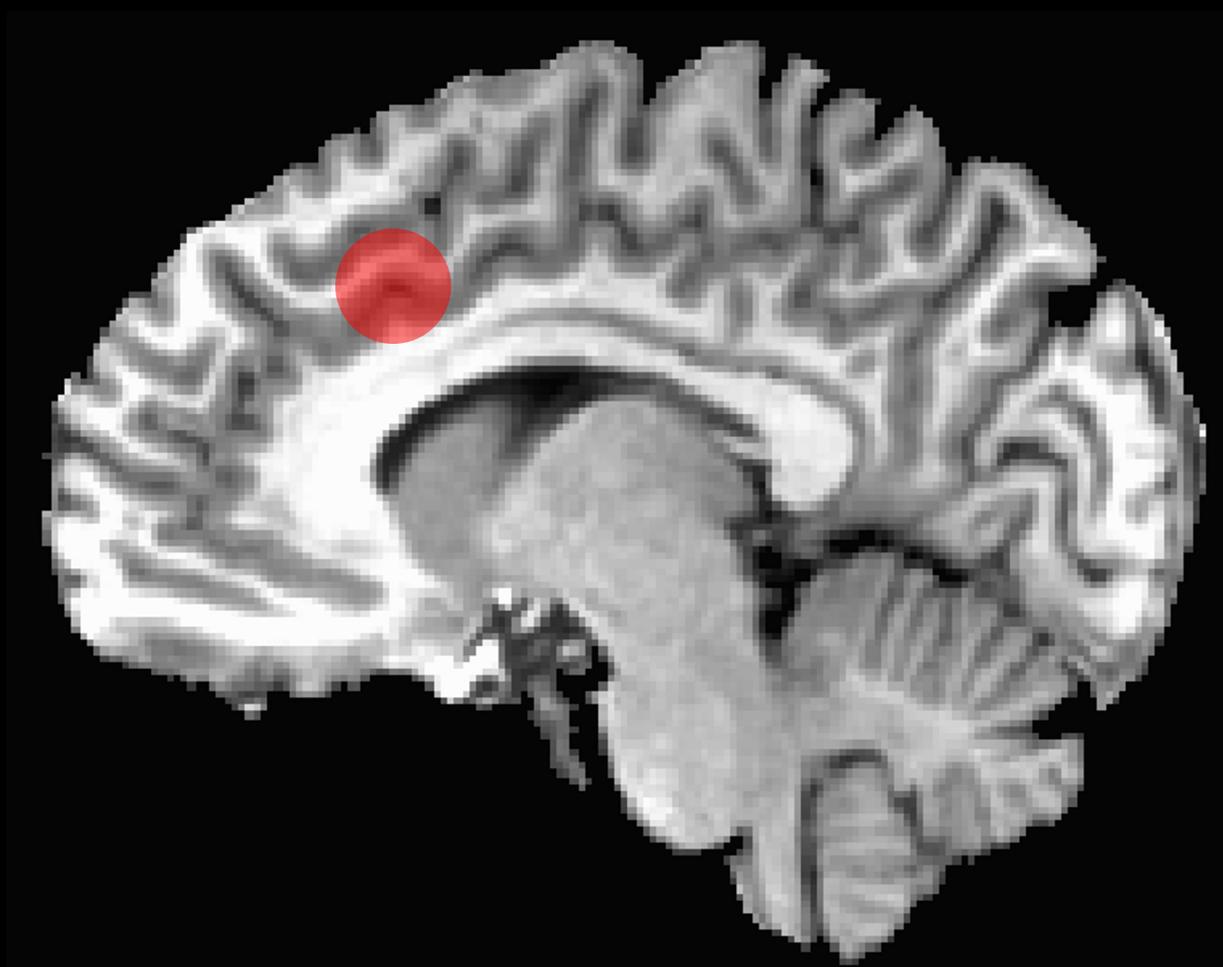


What about this?

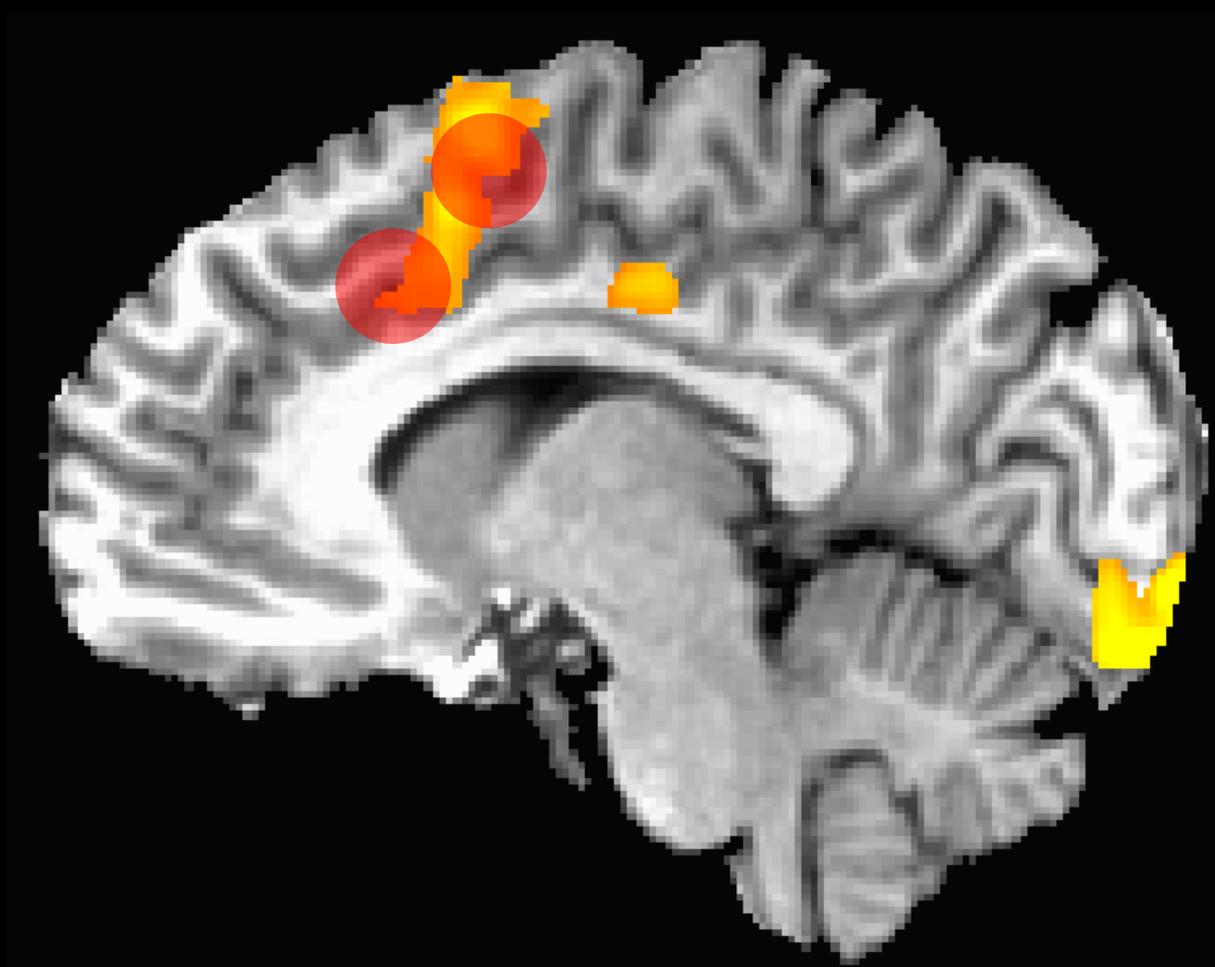


“In this approach, second-level analyses are run for each contrast, consecutively leaving out each subject from the GLM and extracting that subject’s contrast estimates from the resulting ROI.”

What about this?



What about this?



Remember This?

diminished physiological response to stimulation (22).

Examination of regions of interest in the striatum and VMPFC from the gain/loss conjunction analysis (Fig. 3) revealed that these

coefficient of loss aversion (i.e., the ratio of sensitivity to losses versus gains) is highly correlated across risky and riskless contexts (23). Therefore, we surmise that a similar mechanism may contribute to other manifestations of loss aversion.

Fig. 3. Conjunction analysis results. **(A)** Map showing regions with conjointly significant positive gain response and negative loss response ($P < 0.05$, whole-brain corrected, in each individual map) (see also table S1). Red pixels indicate regions showing significant conjunction; green circles highlight clusters included in the respective heatmaps to the right. L, left; R, right. **(B)** Heatmaps were created by averaging parameter estimates versus baseline within each cluster in the conjunction map for each of the 16 cells (of 16 gambles each) in the gain/loss matrix; color coding reflects strength of neural response for each condition, such that dark red represents the strongest activation and dark blue represents the strongest deactivation.

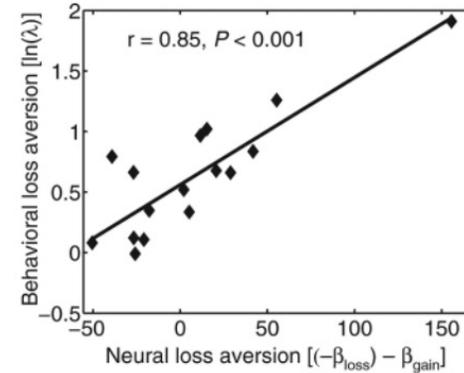
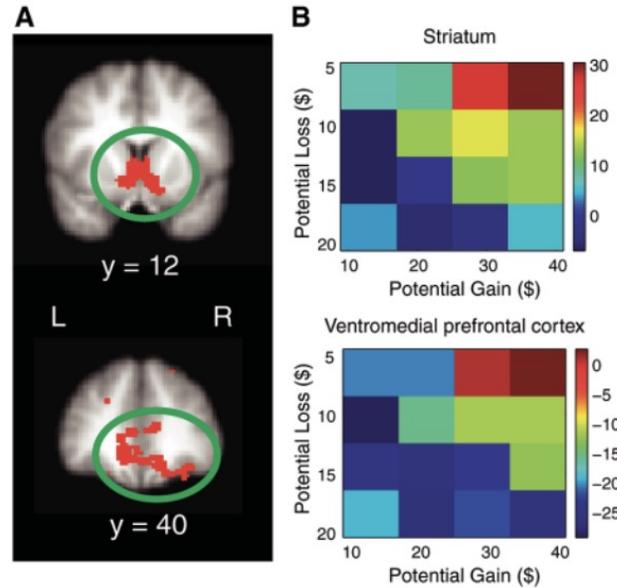


Fig. 4. Scatterplot of correspondence between neural loss aversion and behavioral loss aversion in ventral striatum [Montreal Neurological Institute coordinates (x, y, z) : 3.6, 6.3, 3.9; center of gravity in millimeters]. Regression line and P value were computed with the use of robust regression by iteratively reweighted least squares to prevent the influence of outliers; however, this regression also remained highly significant ($P = 0.004$) when the extreme data point (top right-hand corner) was removed from the analysis. β_{loss} and β_{gain} are the unstandardized regression coefficients for the loss and gain variables, respectively.

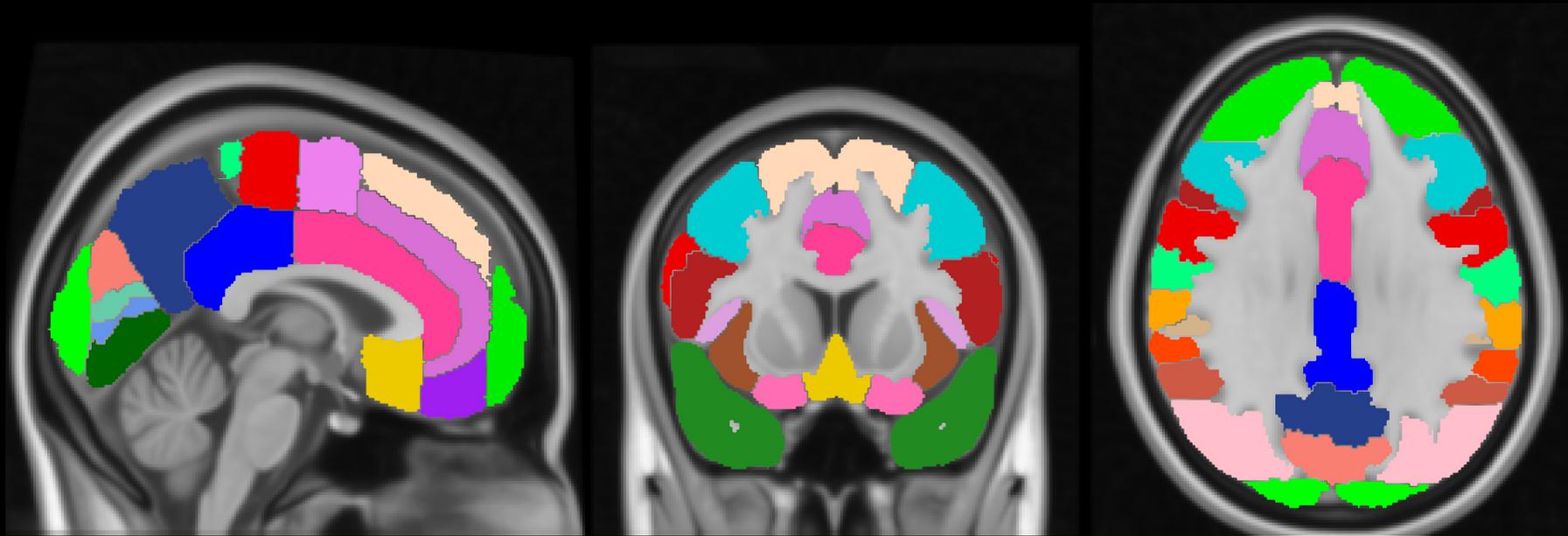
Let's try it!

Demonstration

Resources for Independent Analysis

Atlases

Popular: Harvard-Oxford Atlas, WFU PickAtlas



WFU PickAtlas is a toolbox that needs to be installed

HUMAN ATLAS->TD brodmann areas+

- ..
- brodmann area 1
- brodmann area 2
- brodmann area 3
- brodmann area 4
- brodmann area 5
- brodmann area 6
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- brodmann area 41
- brodmann area 42
- brodmann area 43
- brodmann area 44
- brodmann area 45
- brodmann area 46
- brodmann area 47
- Amygdala
- Anterior Commissure
- Caudate Body
- Caudate Head

Atlas Information

BASIC ADVANCED

ADD ->

MOVE ALL ->>

<- REMOVE SELECTED

<<- REMOVE ALL

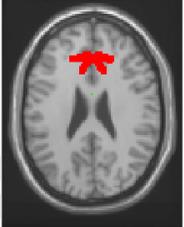
2D 3D

DILATE:

Flip Lock L/R U/D

Left Left + Right Right

Display: Neurologic 49



L R

ITKSnap

Mag

ANALYSIS RESULTS

Write Independent Regions

SAVE MASK

DONE CANCEL

WORKING REGION1

brodmann area 32

	CUBE	46	64	37	GO	ATLAS	SUBREGION	VALUE
MNI		0	0	0	GO	TD brodmann areas+	NA	1000
Tal		0	0	0	GO	TD Lobes	NA	1500

Interrogate Talairach Daemon

Generate Table

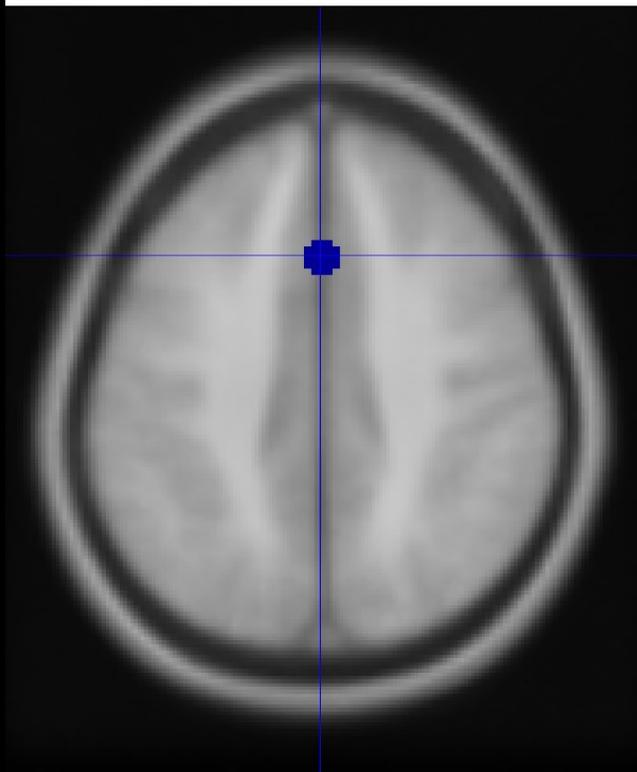
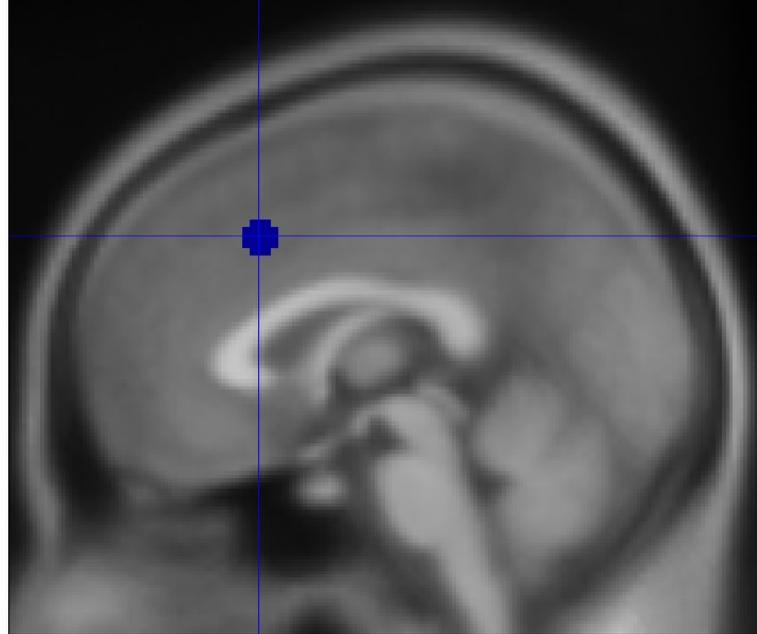
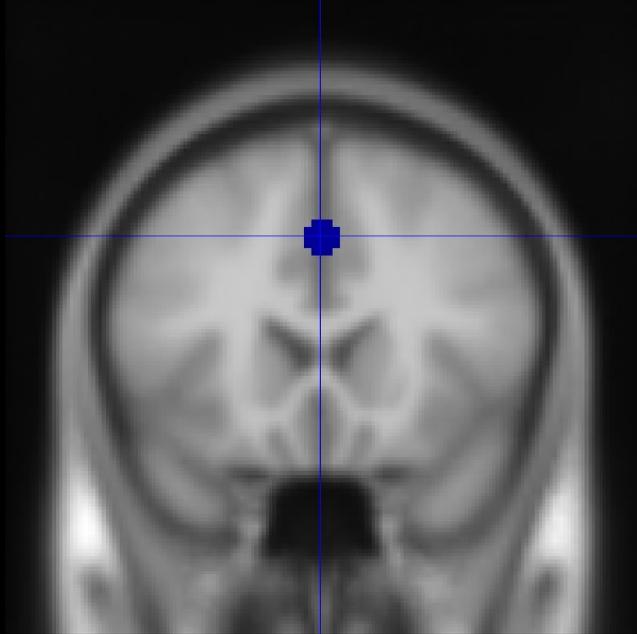
Search Range

Extracting Data from Sphere

Based on coordinates of another study

Table 1. Whole-brain activations at $p < 0.001$ uncorrected, $p < 0.05$ cluster-corrected

Brain region	<i>x</i>	<i>y</i>	<i>z</i>	Z score	Cluster-corrected <i>p</i> value	Cluster size (voxels)
Pain						
Right somatosensory cortex	38	-24	54	5.91	< 0.001	762
Right insula	40	-18	12	5.76	<0.001	902
Left cerebellum	-28	-56	-24	5.43	<0.001	832
Left cingulate gyrus	-2	30	14	5.17	<0.001	788
Right cerebellum	2	-72	-14	4.26	<0.001	524
Right parieto-occipital sulcus	8	-84	40	3.85	<0.001	335



1: dACC_Sphere

Label: dACC_Sphere

Centre of mass: 0 20 40

Volume (mm): 648.00

Max/min X(mm): -4 4

Max/min Y(mm): 16 24

Max/min Z(mm): 36 44

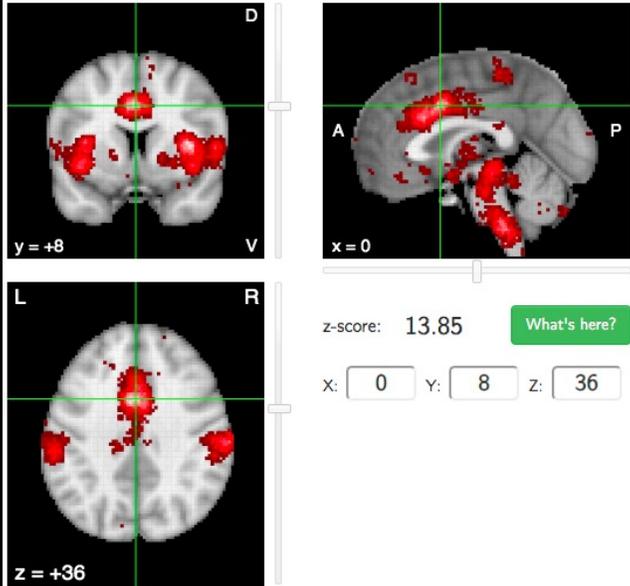
Neurosynth

How it works: Search Terms

pain Search for another term:

An automated meta-analysis of 516 studies

[Maps](#) [Studies](#) [FAQs](#)



z-score: 13.85 What's here?

X: Y: Z:

Layers

<input checked="" type="checkbox"/>	pain: association test	<input type="checkbox"/>	<input type="checkbox"/>
<input checked="" type="checkbox"/>	pain: uniformity test	<input type="checkbox"/>	<input type="checkbox"/>
<input checked="" type="checkbox"/>	anatomical	<input type="checkbox"/>	<input type="checkbox"/>

Color palette:

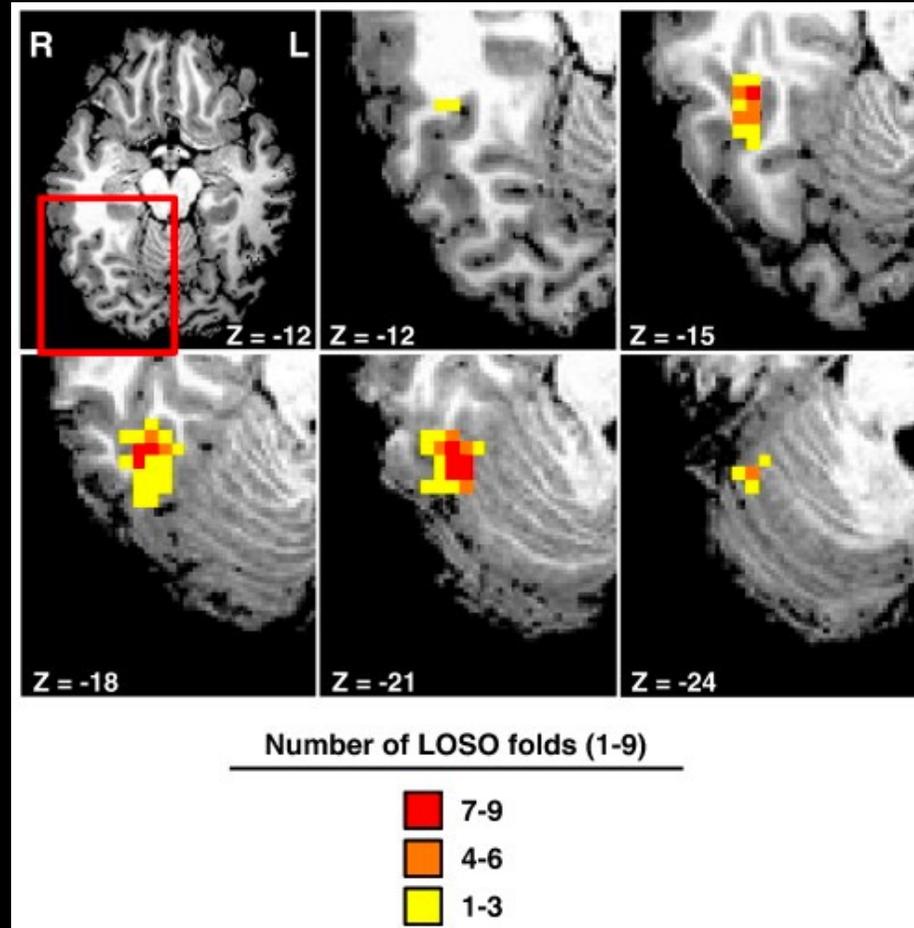
Positive/Negative:

Thresholds:

Opacity:

Crosshairs
 Pan/zoom
 Labels

Cross-Validation



Esterman et al., 2010

Demonstration

Biased Analyses: What to do about it?

Choose an independent analysis

Know the reasons behind choosing it

**An “independent” analysis does not guarantee
non-biased results!**

Cluster Failure: A Discussion of Eklund et al. (2016)

PNAS

...we found that the most common software packages for fMRI...can result in false-positive rates of up to 70%. These results question the validity of some *40,000 fMRI studies*. [Emphasis added.]



The
Economist

When science goes wrong (I)

Computer says: oops

Two studies, one on neuroscience and one on palaeoclimatology, cast doubt on established results. First, neuroscience and the reliability of brain scanning

Circular analysis in systems neuroscience: the dangers of double dipping

Nikolaus Kriegeskorte, W Kyle Simmons, Patrick S F Bellgowan & Chris I Baker

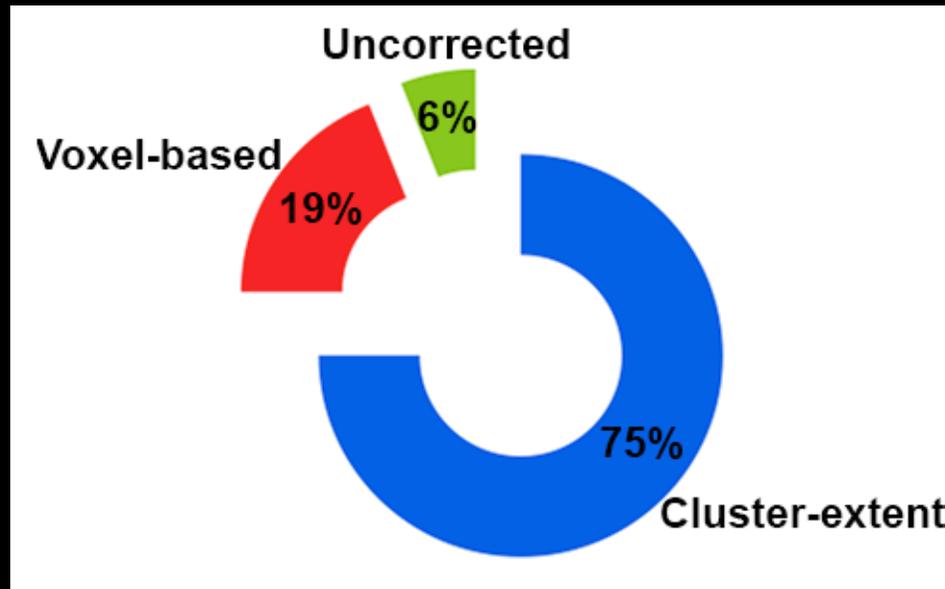
Erroneous analyses of interactions in neuroscience: a problem of significance

Sander Nieuwenhuis^{1,2}, Birte U Forstmann³ & Eric-Jan Wagenmakers³

Puzzlingly High Correlations in fMRI Studies of Emotion, Personality, and Social Cognition¹

Edward Vul,¹ Christine Harris,² Piotr Winkielman,² & Harold Pashler²

¹Massachusetts Institute of Technology and ²University of California, San Diego



Adapted from Woo et al. (2014)

Is this true?

How big of a problem is this?

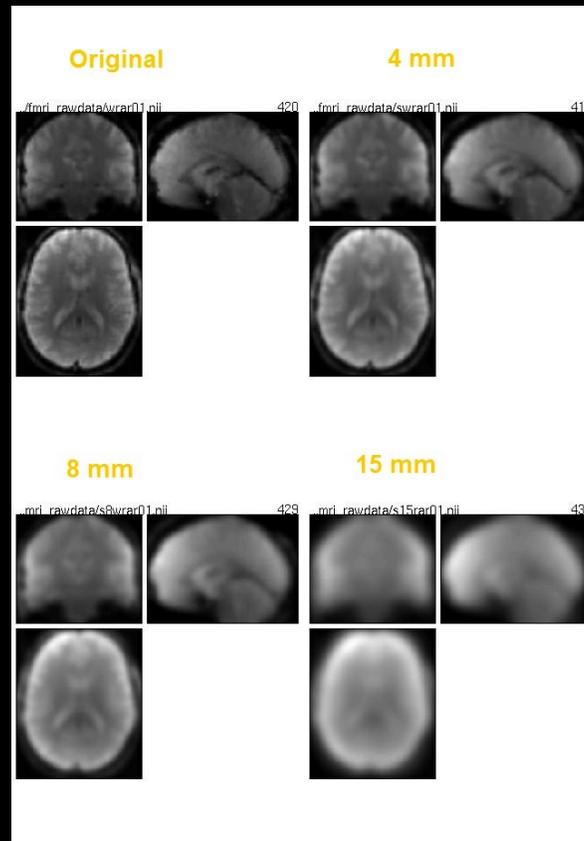
What should we do about it?

1. Review of cluster thresholding

2. Problems with cluster thresholding

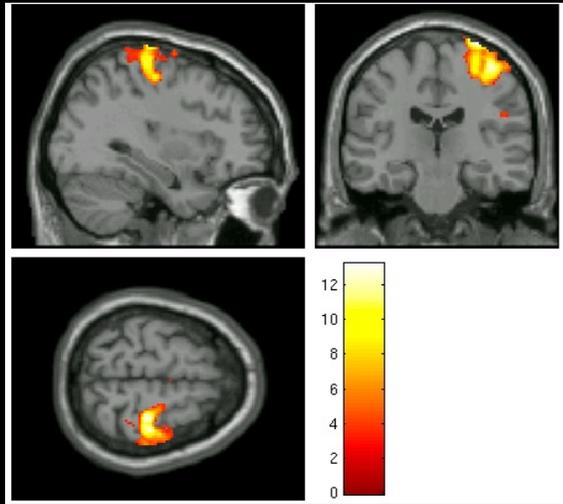
3. How to address these problems

Smoothing increases spatial correlation

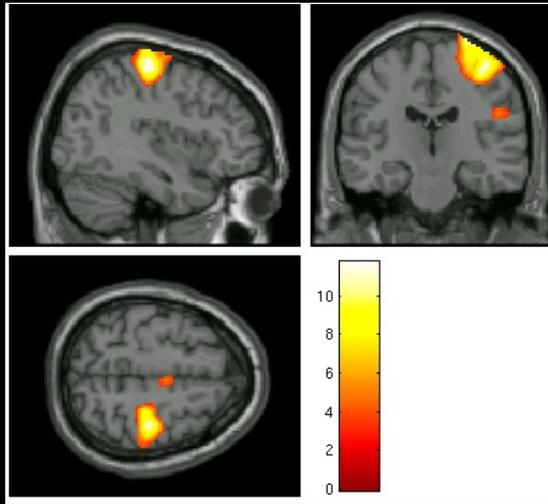


Smoothing increases cluster size

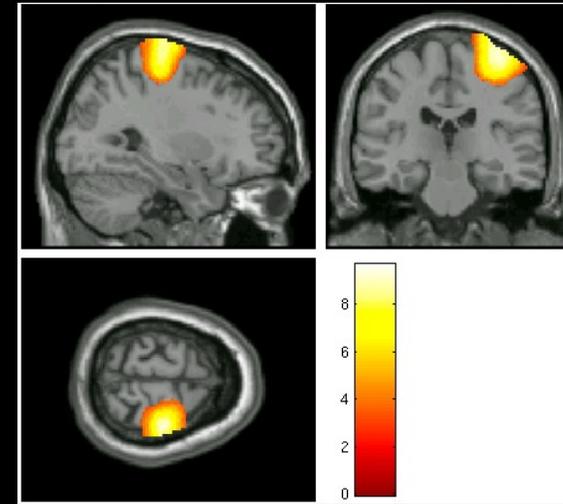
4mm



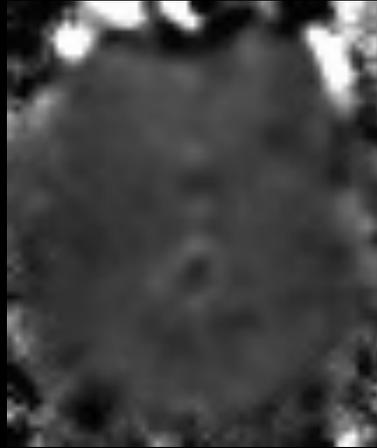
8mm



15mm



Smoothness is estimated from the residuals



3dFWHMx -mask mask.hdr ResMS.hdr

++ 3dFWHMx: AFNI version=AFNI_16.1.28 (Jun 30 2016) [64-bit]

++ Authored by: The Bob

++ Number of voxels in mask = 174611

++ start FWHM calculations

+ FWHM done (0.00 CPU s thus far)

11.8179 12.9726 12.9531

These smoothness estimates form cluster thresholds

```
3dClustSim -mask mask.hdr -fwhm 12.5 -athr 0.05 -pthr 0.01
```

```
++ 3dClustSim: AFNI version=AFNI_16.1.28 (Jun 30 2016) [64-bit]
++ 174611 voxels in mask (19.34% of total)
++ Padding by 10 x 10 x 10 slices to allow for edge effects of blurring
++ Startup clock time = 0.0 s
++ Using 8 OpenMP threads
# CLUSTER SIZE THRESHOLD(pthr,alpha) in Voxels
# -NN 3 | alpha = Prob(Cluster >= given size)
```

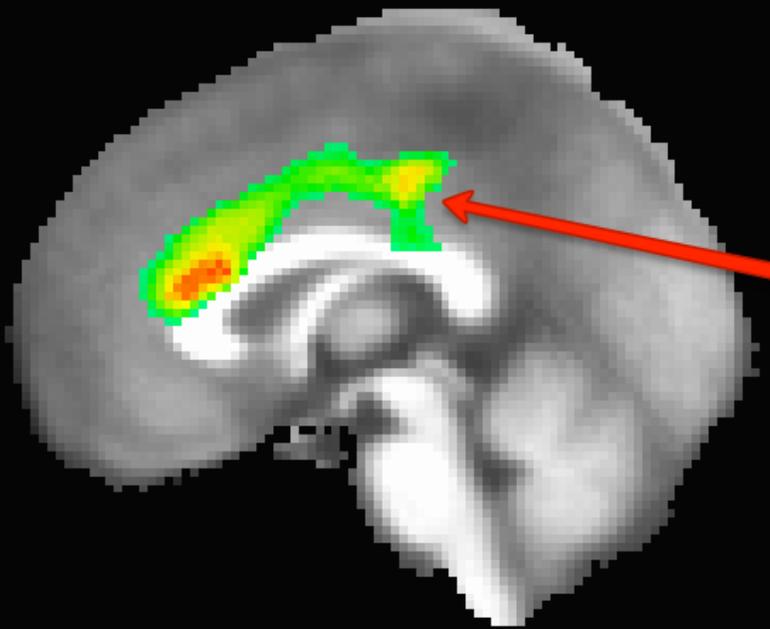
```
# pthr | .05000
```

```
# ----- | -----
```

```
0.010000 667.8
```

i.e., A cluster size of 668 voxels or more is significant

Determine whether clusters pass threshold



Voxels survived clustering = 11234
Voxels edited out = 1776
NN clustering level = 1 [faces touch]

Use internal mask from 3dClustSim | linkRbrain type Tasks

#3 | XYZ Peak | 3dclust SaveTabl Clust SaveMsk WamI | Done

###:	__Size__	__X__	__Y__	__Z__	Alpha					
<input type="checkbox"/> 1:	5496 vox	+28.0	+56.0	-24.0	Jump	Flash	Plot	Save	Write	N/Csim
<input type="checkbox"/> 2:	1702 vox	-40.0	+18.0	+12.0	Jump	Flash	Plot	Save	Write	N/Csim
<input checked="" type="checkbox"/> 3:	1572 vox	+2.0	-30.0	+14.0	Jump	Flash	Plot	Save	Write	N/Csim
<input type="checkbox"/> 4:	1239 vox	-38.0	+24.0	+54.0	Jump	Flash	Plot	Save	Write	N/Csim
<input type="checkbox"/> 5:	1225 vox	+38.0	+20.0	+12.0	Jump	Flash	Plot	Save	Write	N/Csim

Greater than 668?

Problems with Cluster Thresholding

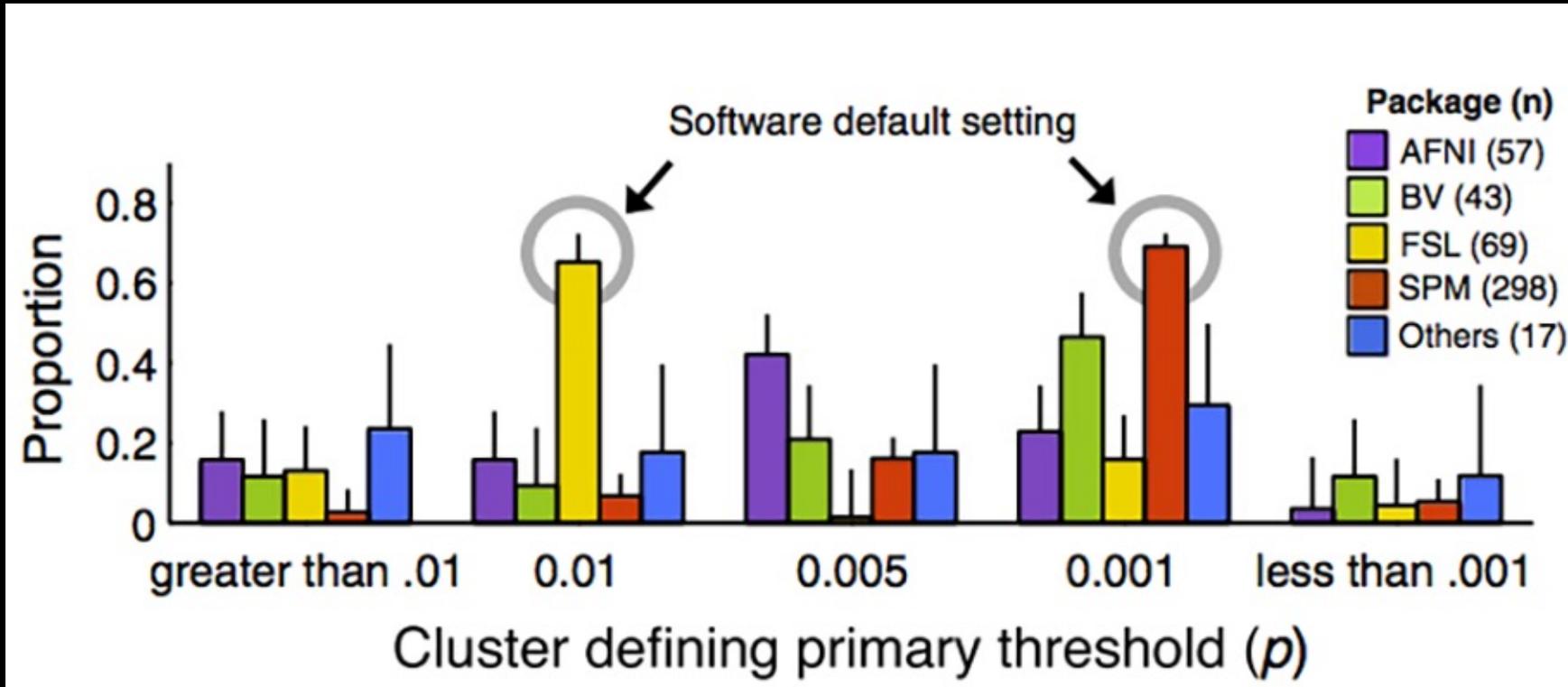
1. Using default thresholds

2. Clusters span multiple areas

3. Cluster simulations violate parametric assumptions

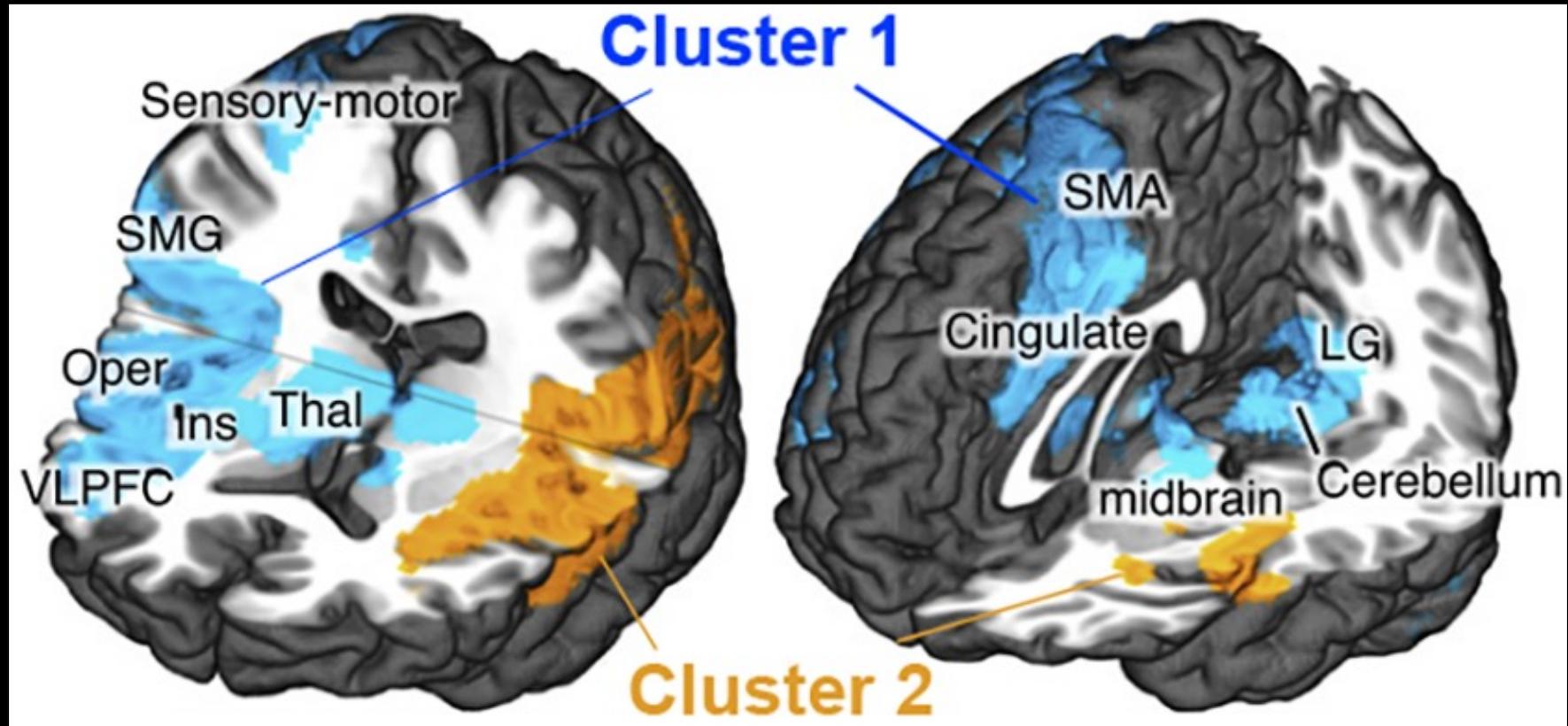
Problem #1

Using default thresholds



Problem #2

Clusters span multiple areas



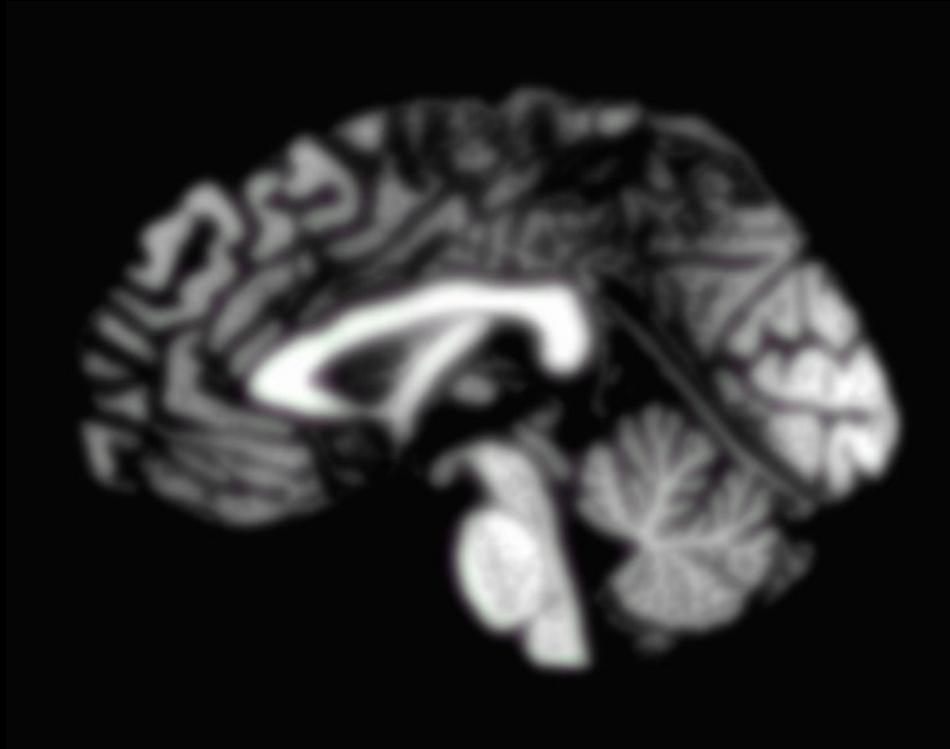
Problem #3

Violation of assumptions

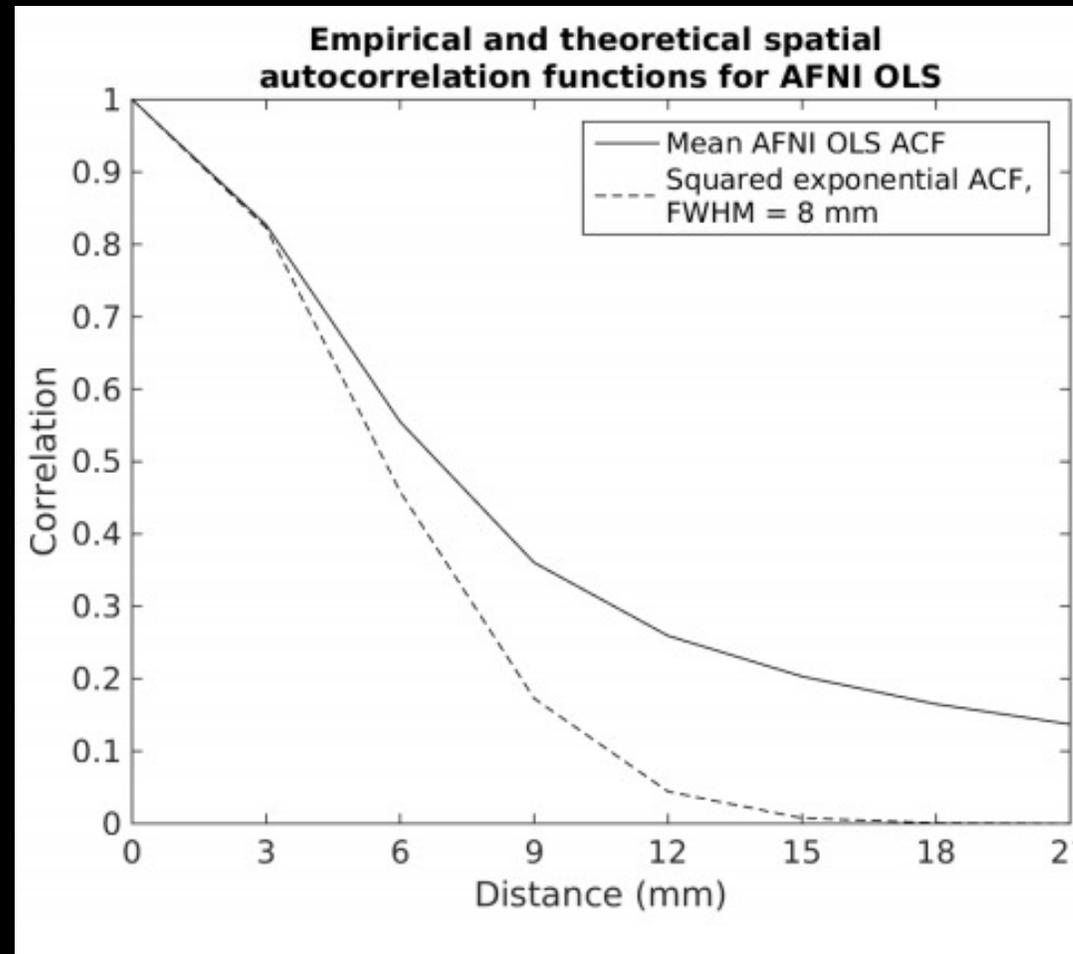
Cluster thresholding methods assume that:

- 1) Spatial smoothness is constant over the entire brain; and**
- 2) Spatial autocorrelation is normally distributed**

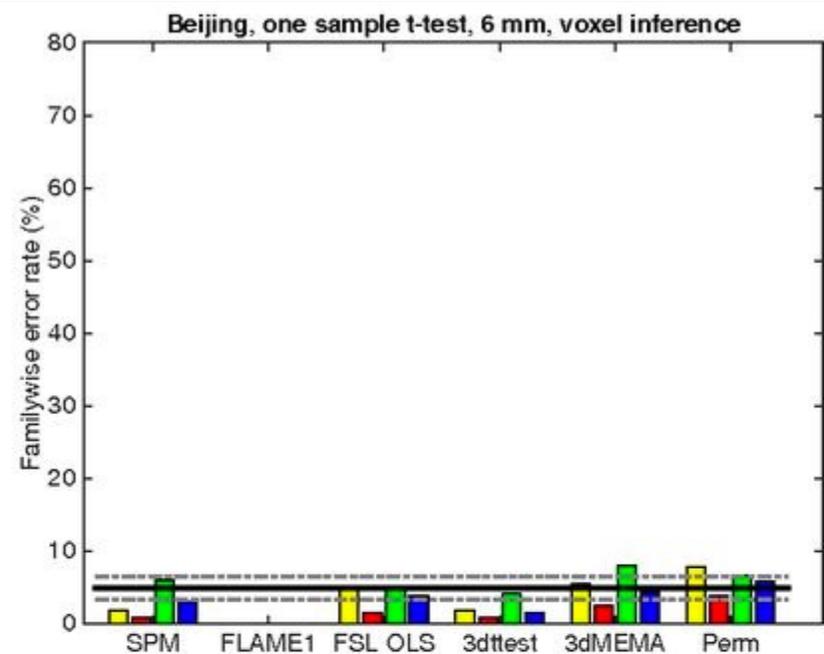
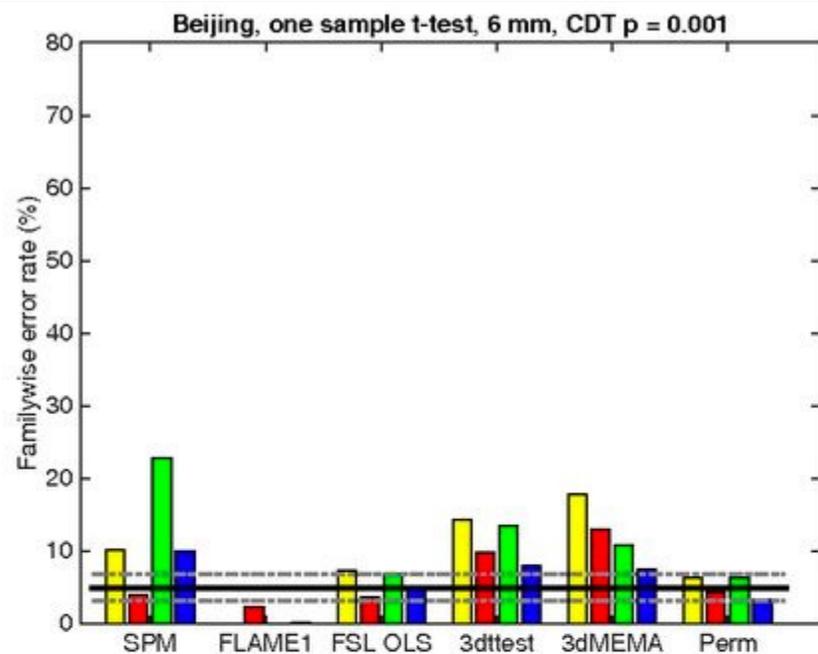
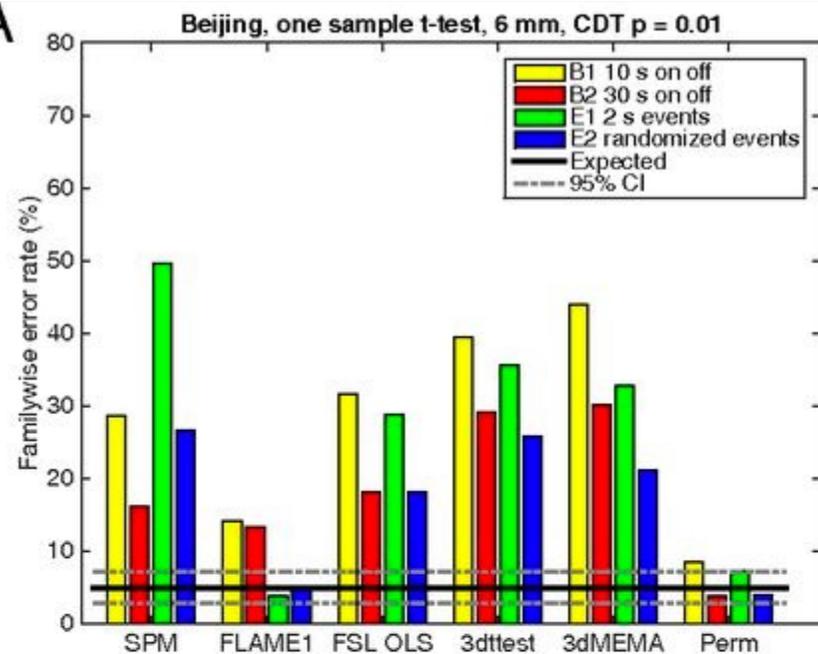
1. Smoothness varies over the brain



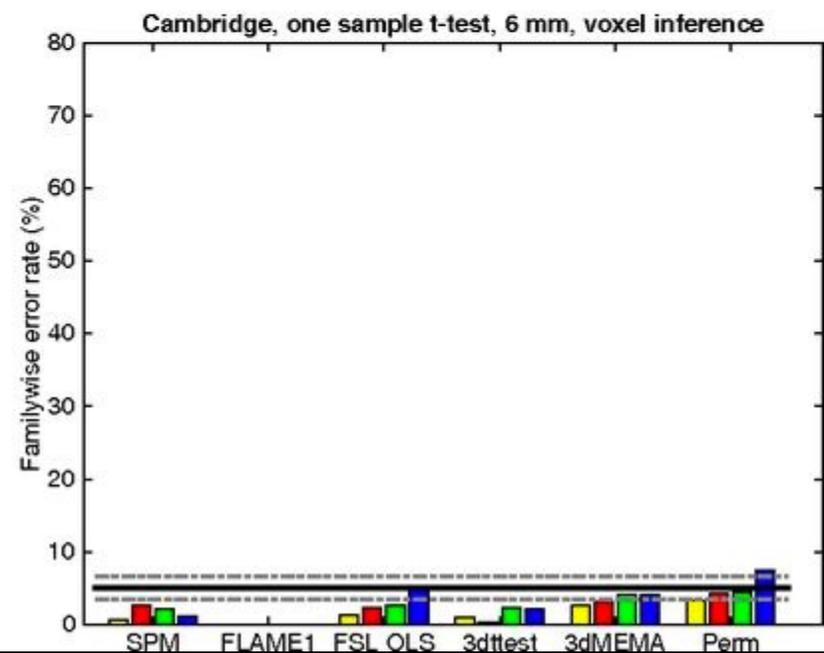
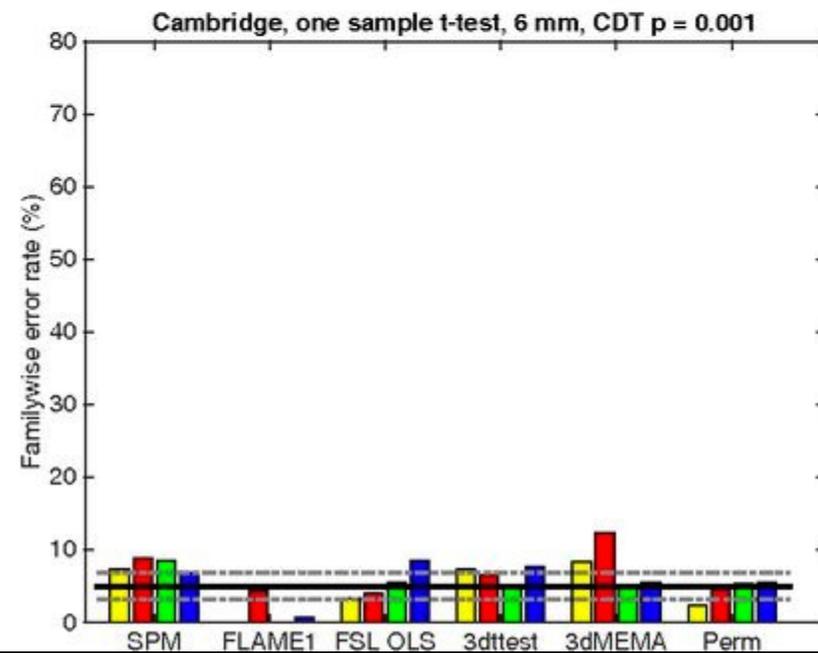
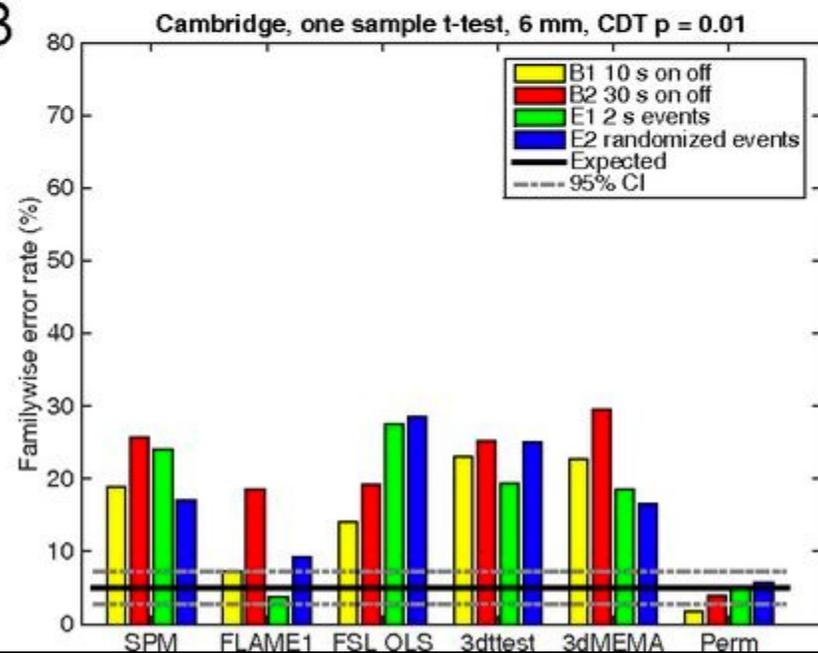
2. Spatial autocorrelation is best modeled as a mixture of Gaussian and exponential distributions



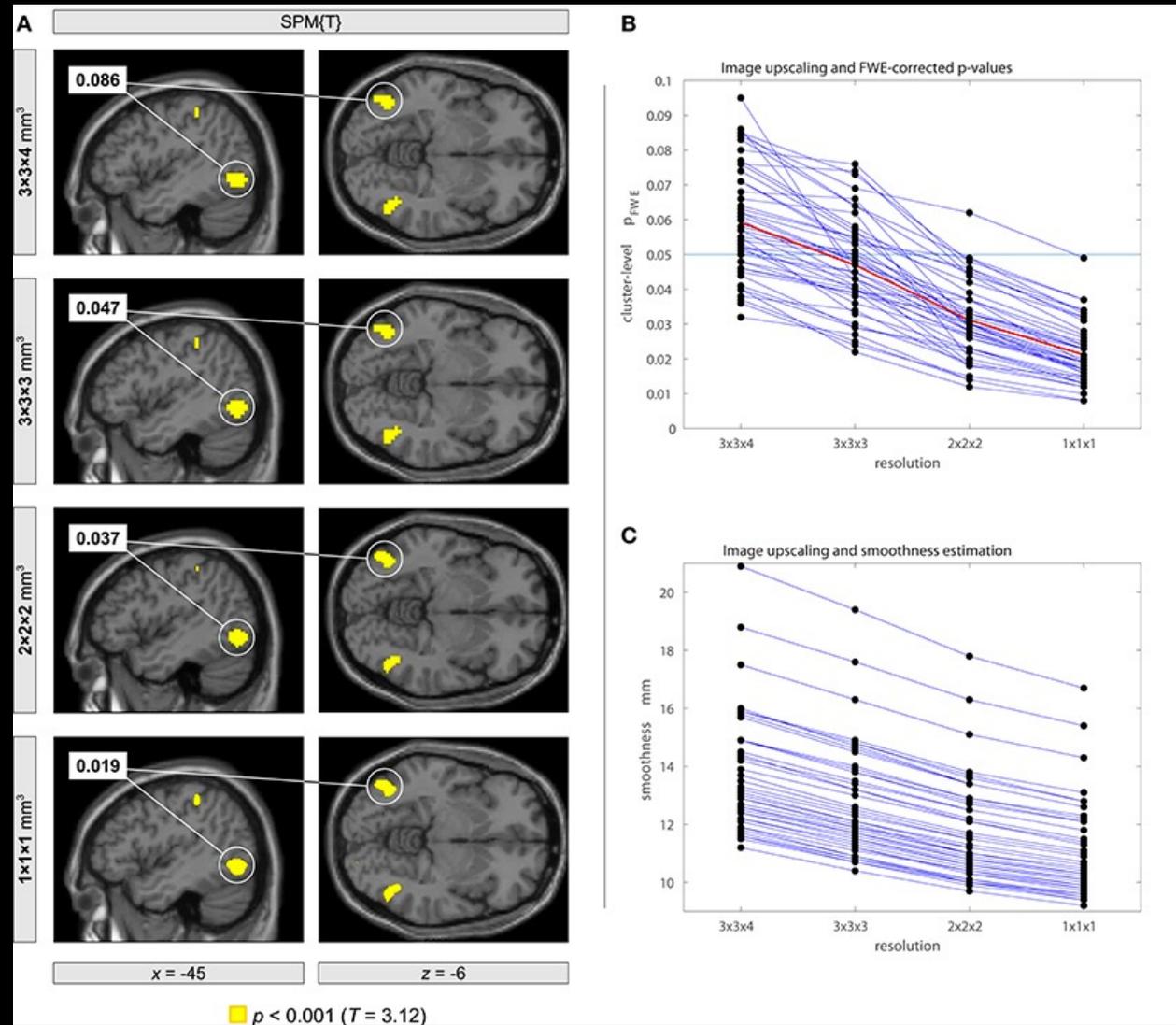
A



B



Aside: Does Voxel Size affect FPR?



Should we trust cluster corrected results from SPM?

Unclear how SPM has addressed this in latest updates

FSL seems unaffected with default settings

AFNI has made attempts to address this, as we will see

$p=0.010$

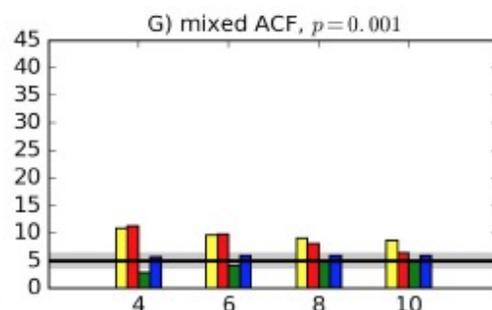
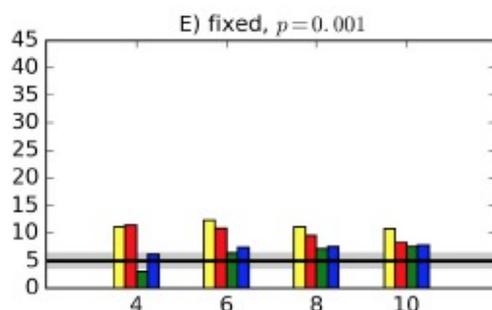
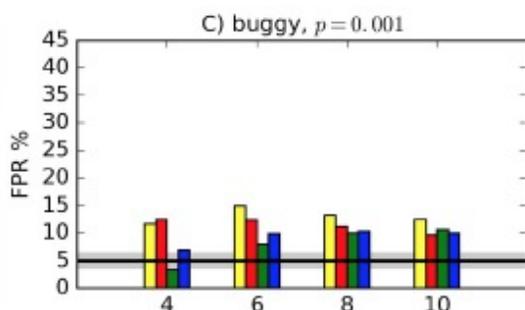
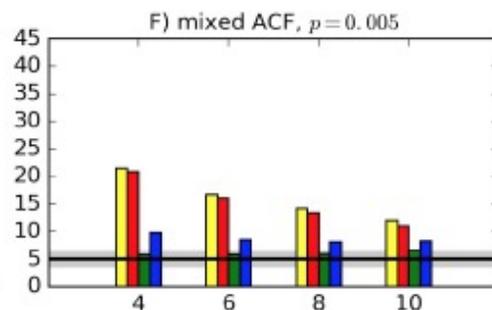
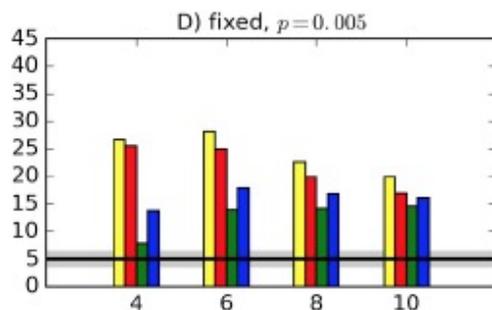
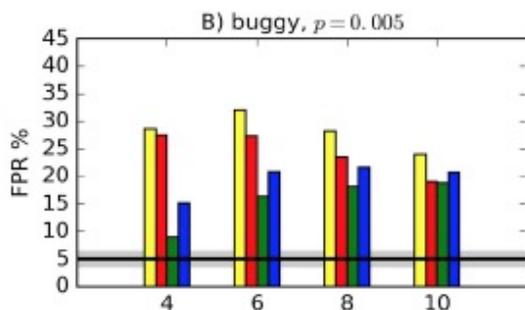
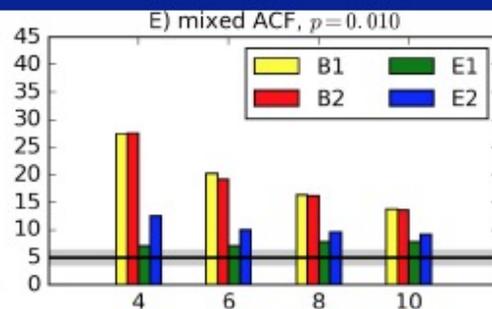
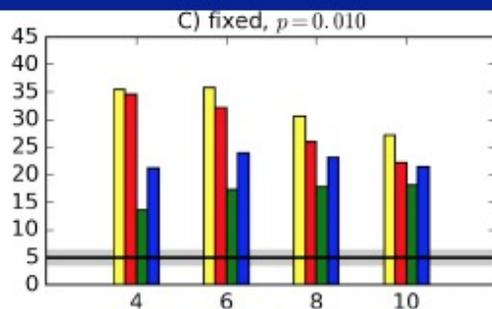
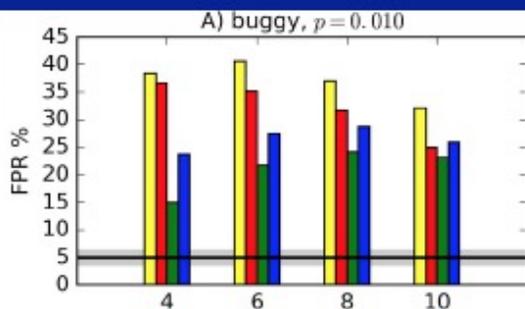
$p=0.005$

$p=0.001$

Pre-bug fix

Post-bug fix

Mixed-model ACF



smoothing value (mm)

smoothing value (mm)

smoothing value (mm)

Solutions

1) Use voxel-wise thresholds;

2) Use more conservative cluster thresholds; or

3) Use nonparametric methods (e.g., permutations)

Non-parametric tests

FSL's randomise (requires creating a design matrix)

SPM's SnPM (separate toolbox)

Non-parametric tests

No assumption of the distribution the data were drawn from

The data itself is used to construct a new distribution

**Example: We have a positively-skewed distribution,
and still want to use p-values**

Non-parametric tests

A	B	A	B	A	B
103.00	90.48	99.93	87.83	99.76	96.06

Experiment with two conditions: A=StimOn, B=StimOff

Null Hypothesis: Labels are arbitrary

Non-parametric tests

Construct a set of all possible relabelings

AAABBB

ABABAB

BAAABB

BABBAA

AABABB

ABABBA

BAABAB

BBAAAB

AABBAB

ABBAAB

BAABBA

BBAABA

AABBBA

ABBABA

BABAAB

BBABAA

ABAABB

ABBBAA

BABABA

BBBAAA

Non-parametric tests

Construct a set of all possible relabelings

AAABBB	4.82	ABABAB	9.45	BAAABB	-1.48	BABBAA	-6.86
AABABB	-3.25	ABABBA	6.97	BAABAB	1.10	BBAAAB	3.15
AABBAB	-0.67	ABBAAB	1.38	BAABBA	-1.38	BBAABA	0.67
AABBBA	-3.15	ABBABA	-1.10	BABAAB	-6.97	BBABAA	3.25
ABAABB	6.86	ABBBAA	1.48	BABABA	-9.45	BBBAAA	-4.82

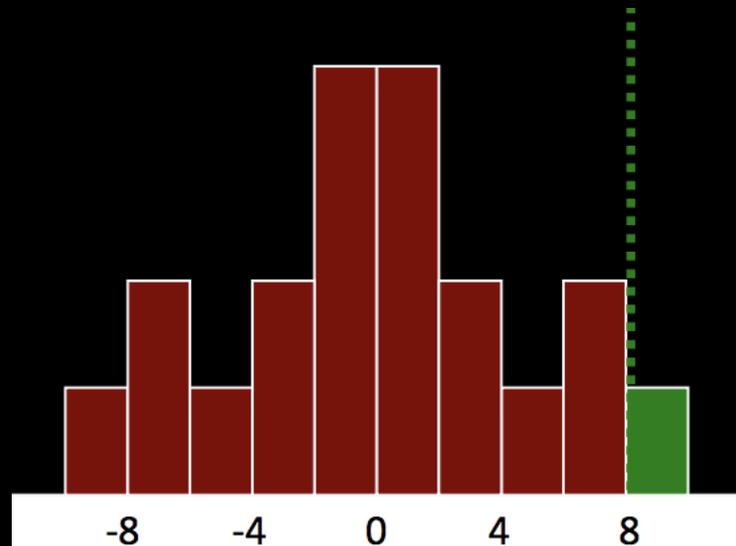
Non-parametric tests

Construct a set of all possible relabelings

AAABBB	4.82	ABABAB	9.45	BAAABB	-1.48	BABBAA	-6.86
AABABB	-3.25	ABABBA	6.97	BAABAB	1.10	BBAAAB	3.15
AABBAB	-0.67	ABBAAB	1.38	BAABBA	-1.38	BBAABA	0.67
AABBBA	-3.15	ABBABA	-1.10	BABAAB	-6.97	BBABAA	3.25
ABAABB	6.86	ABBBAA	1.48	BABABA	-9.45	BBBAAA	-4.82

Non-parametric tests

Construct a set of all possible relabelings



SnPM

The screenshot shows the 'Batch Editor' window with the 'SPM' menu open. The 'Tools' sub-menu is selected, and the 'SnPM' option is highlighted. The 'Specify' submenu is also open, displaying a list of statistical tests:

- SingleSub: Two Sample T test; 2 conditions, replications
- SingleSub: Simple Regression (correlation); single covariate of interest
- MultiSub: One Sample T test on diffs/contrasts**
- MultiSub: Simple Regression; 1 covariate of interest
- MultiSub: Paired T test; 2 conditions, 1 scan per condition
- Within Subject ANOVA, k diffs/contrasts per subject
- 2 Groups: Two Sample T test; 1 scan per subject
- 2 Groups: Test diff of response; 2 conditions, 1 scan per condition
- Between group ANOVA; 1 scan per subject

The background interface includes a 'Module List' pane with 'No Modules in' and a 'MATLAB Batch System' section with 'Matlabbatch User Interface' and '* Menu and Toolbar'.

SnPM

The screenshot shows the SnPM Batch Editor window. The title bar reads "Batch Editor". The menu bar includes "File", "Edit", "View", "SPM", and "BasicIO". The main interface is divided into several sections:

- Module List:** A list on the left with "MultiSub: One Sample T test on" selected.
- Current Module:** "MultiSub: One Sample T test on diffs/contrasts".
- Help:** "MultiSub: One Sample T test on diffs/contrasts".
- Analysis Directory:** "...ts\2x2\Face-Hand\nonParametric".
- Images to analyze:** 10 files.
- Covariates:** (empty).
- Number of Permutations:** 5000.
- Variance smoothing:** [0 0 0].
- Memory usage:** High.
- Cluster inference:** . None.
- Masking:** . Threshold masking.
- Implicit Mask:** Yes.
- Explicit Mask:** . None.
- Global calculation:** . Omit.
- Global normalisation:** . Overall grand mean scaling.
- Normalisation:** . No.

Current Item: Images to analyze

- C:\fMRI_Course\Data\SPM_Labs\Subjects\sub01\results\precooked\2x2\con_0002.img,1
- C:\fMRI_Course\Data\SPM_Labs\Subjects\sub02\results\precooked\2x2\con_0002.img,1
- C:\fMRI_Course\Data\SPM_Labs\Subjects\sub03\results\precooked\2x2\con_0002.img,1
- C:\fMRI_Course\Data\SPM_Labs\Subjects\sub04\results\precooked\2x2\con_0002.img,1
- C:\fMRI_Course\Data\SPM_Labs\Subjects\sub05\results\precooked\2x2\con_0002.img,1
- C:\fMRI_Course\Data\SPM_Labs\Subjects\sub06\results\precooked\2x2\con_0002.img,1
- C:\fMRI_Course\Data\SPM_Labs\Subjects\sub07\results\precooked\2x2\con_0002.img,1

At the bottom, there is a "Specify..." button and a section titled "Images to analyze" with the text "Model will be fit to these images."

SnPM

Batch Editor

File Edit View SPM BasicIO

Module List

- MultiSub: One Sample T test on diffs.
- Compute
- Inference

Current Module: Inference

Help on: Inference

SnPM.mat results file DEP Compute: SnPM.mat results file

Type of Thresholding

- . Cluster-Level Inference
- . . Cluster size
- . . . Cluster-Forming Threshold 0.001
- . . . Significance Level
- . . . FWE Corrected 0.05

Display positive or negative effects? Positives

Write thresholded/filtered statistic image?

- . Image name to write SnPM_filtered
- Results displayed: Standard (MIP & table)

Current Item: SnPM.mat results file

Reference from

Compute: SnPM.mat results file

Specify...

Dependency

SnPM.mat results file

Select a SnPM.mat results file.

Non-Parametric Analyses: Pros and Cons

Pros: Tends to be accurate for multiple comparisons

No assumption about the shape of the distribution

Cons: Conservative for small sample sizes

Assumes exchangeability of data

3dttest++

Using the `-Clustsim` option will generate a table for each cluster-wise p-threshold

Which threshold you use is up to you; there is nothing taboo about using 0.01 or 0.05

AFNI Distributions since 2019 will use mixed ACF

AFNI Cluster Results [A]

Voxels survived clustering = 3748
 Voxels edited out = 1207
 # Positive clusters = 12
 # Negative clusters = 0
 NN clustering level = 2 [edges touch]

Alpha -> Cluster thresh: 0.10->71 : 0.05->116 : 0.01->296

Use internal mask from 3dClustSim

#5 XYZ Peak 3dclust SaveTabl Clust SaveMsk WanI Done

###	Size	X	Y	Z	Alpha
1	922 vox	+39.0	+87.0	-9.0	<<0.01
2	716 vox	-33.0	+72.0	+48.0	<<0.01
3	532 vox	-42.0	-24.0	+9.0	< 0.01
4	404 vox	-45.0	+66.0	-6.0	< 0.01
5	299 vox	-12.0	-24.0	+39.0	< 0.01
6	280 vox	+0.0	+60.0	-36.0	< 0.02

AFNI News AFNI Forum
 AFNI Tips Prog Helps

Original View
 AC-PC Aligned
 Talairach View

Define Overlay ->
 See Overlay

Define Datamode ->

DataDir Switch Read

UnderLay EditEnv
 OverLay NIIM+PO

Control Surface

Thr A B 0Lay Edit 0Lay InstaCorr

*Clusterize Setup ICorr
 Clear Rpt *NOT Ready*

ULay #0 SKoff
 0Lay #1 Inc-Con_Zscr
 Thr #1 Inc-Con_Zscr

ULay 1570
 0Lay -4.00943: 5.463681
 Thr -4.09143: 5.463681

See TT Atlas Regions

ULay = 90
 0Lay = 3.286194
 Thr = 3.286194

p=.0100*
 q=.1094

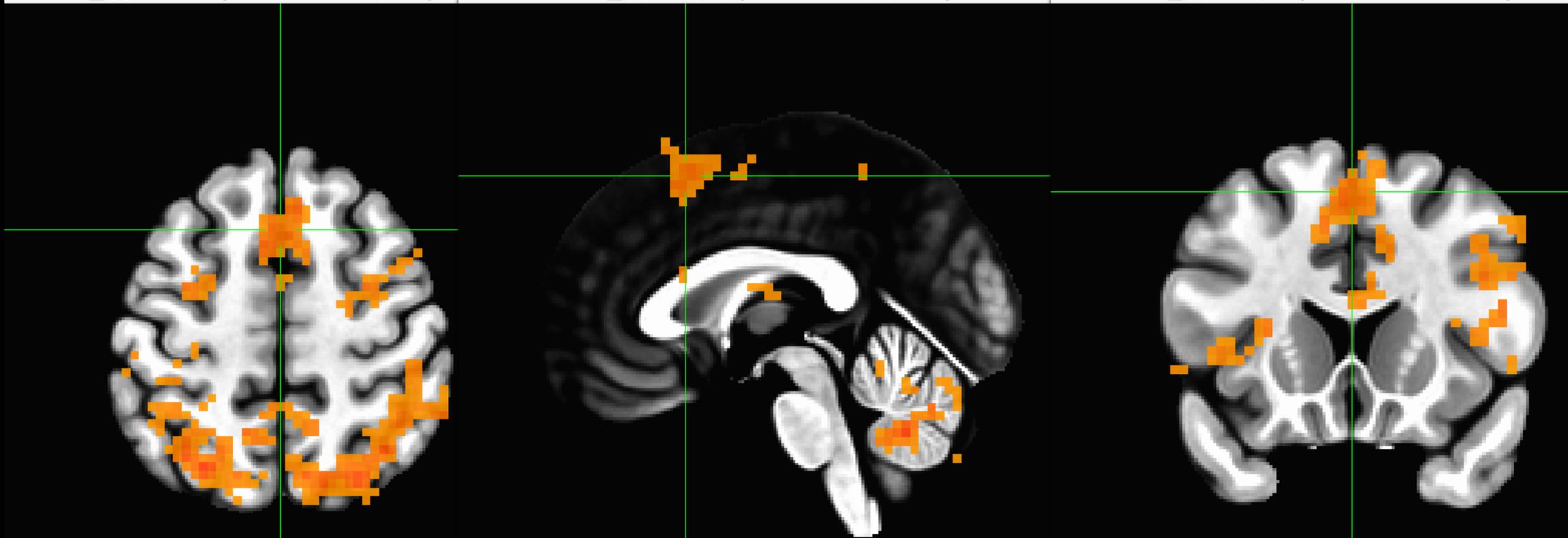
10^1

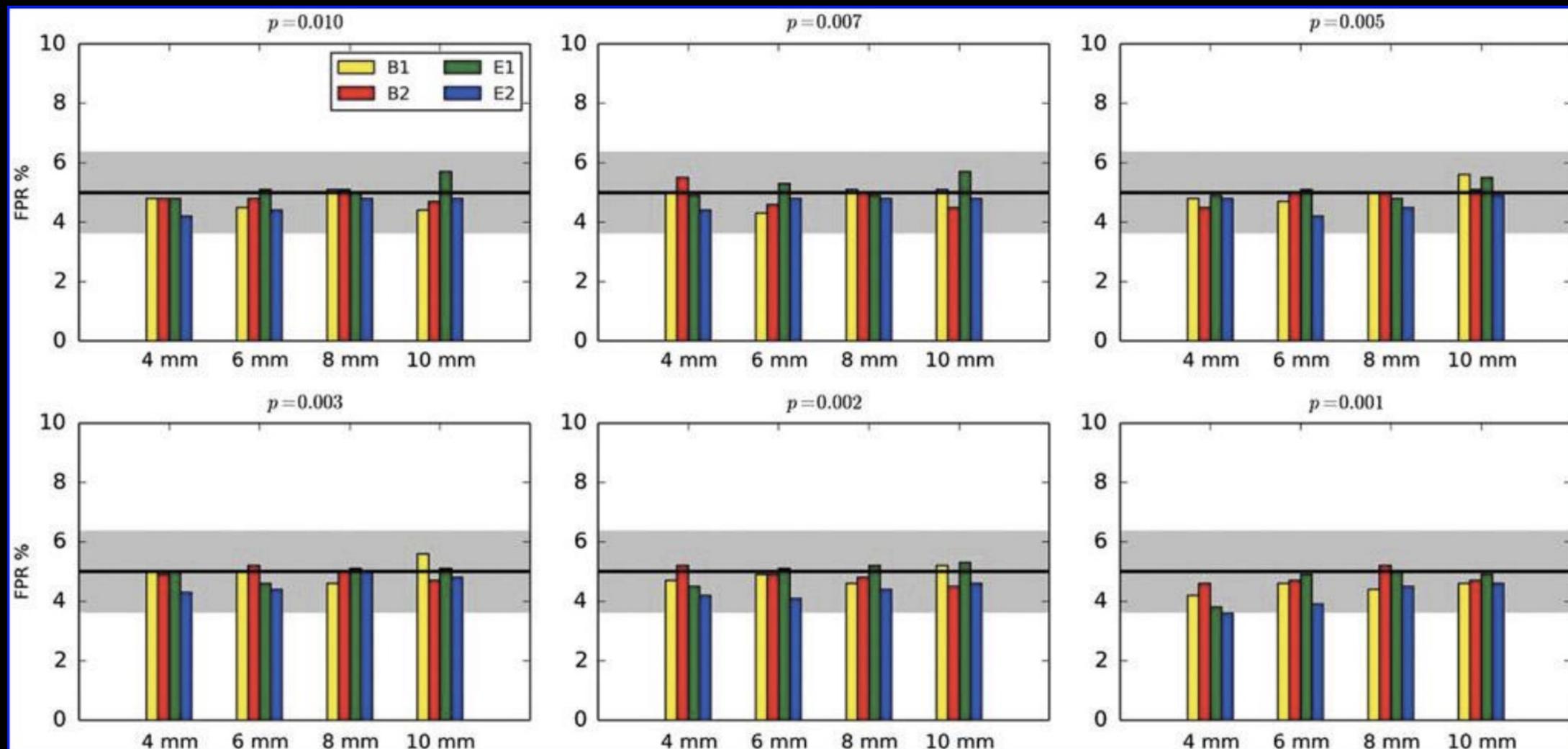
Order: RAI=DICOM
 x = 0.000 mm
 y = -17.000 mm [A]
 z = 54.000 mm [S]

Xhairs Multi X+
 Color green
 Gap 5 Wrap
 Index 1

Axial Image Graph
 Sagittal Image Graph
 Coronal Image Graph

New Etc->
 BHelp done





3dClustSim Demo

Problems with 3dClustSim

Still a risk of false negatives

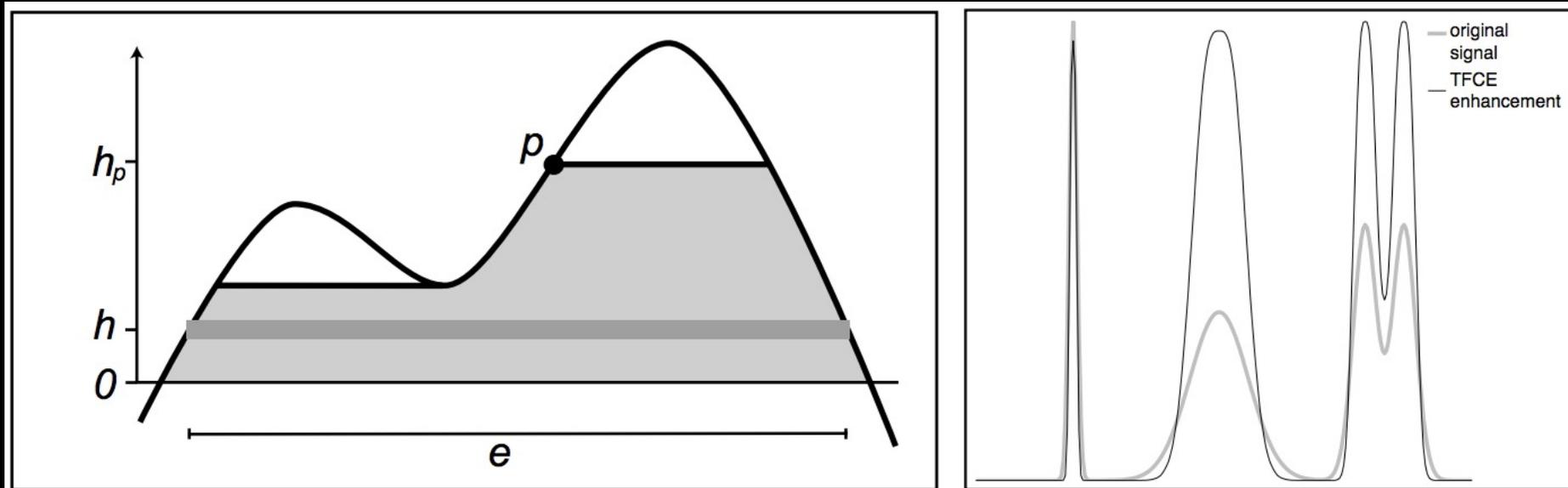
Uses arbitrary threshold set by user

Threshold-Free Cluster Enhancement (TFCE)

What if we didn't have to set a cluster-forming threshold?

Calculate the area under the curve

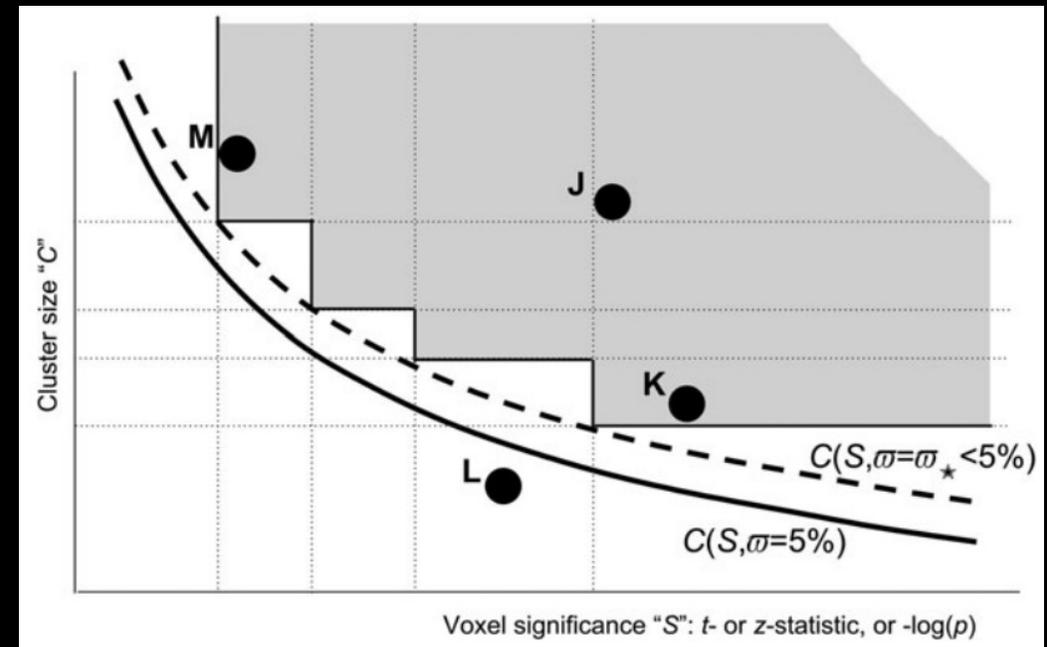
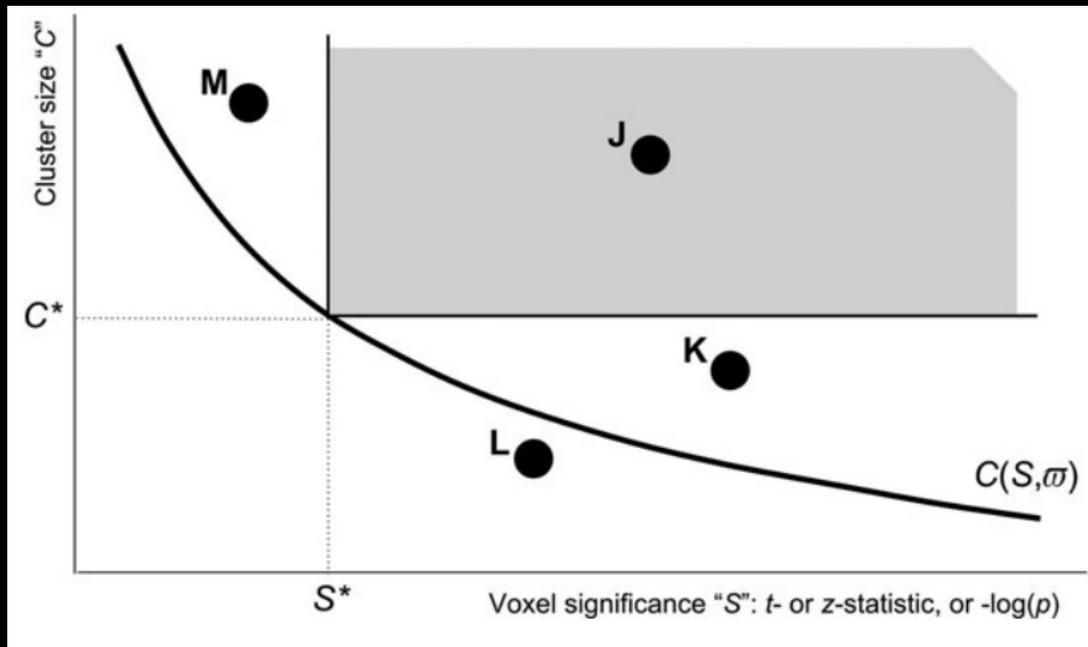
Similar to AFNI's Equitable Thresholding and Clustering (ETAC)



Threshold-Free Cluster Enhancement (TFCE)

No parametric assumptions about shaped of smoothness

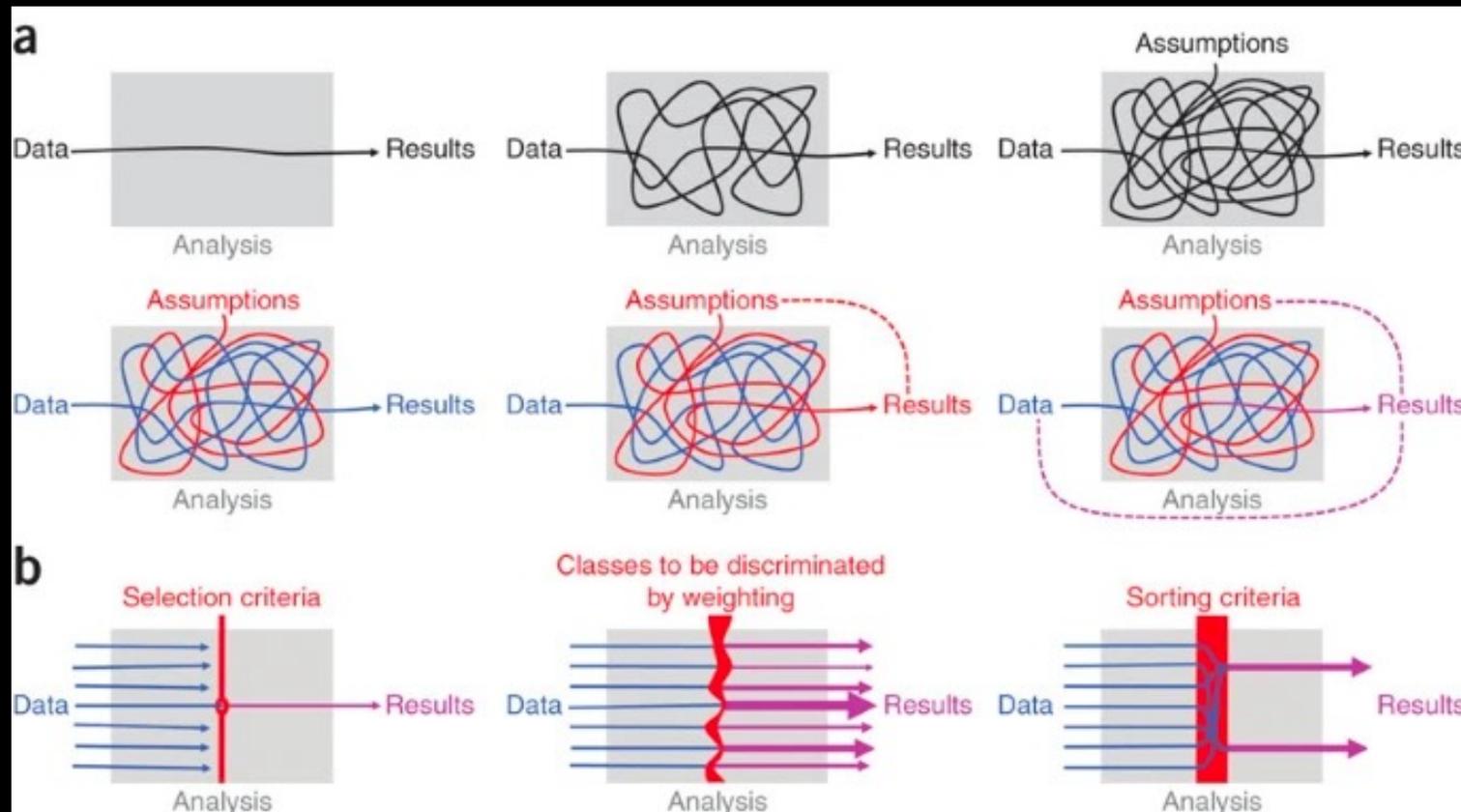
Balances the extent and height thresholds at constant FPR



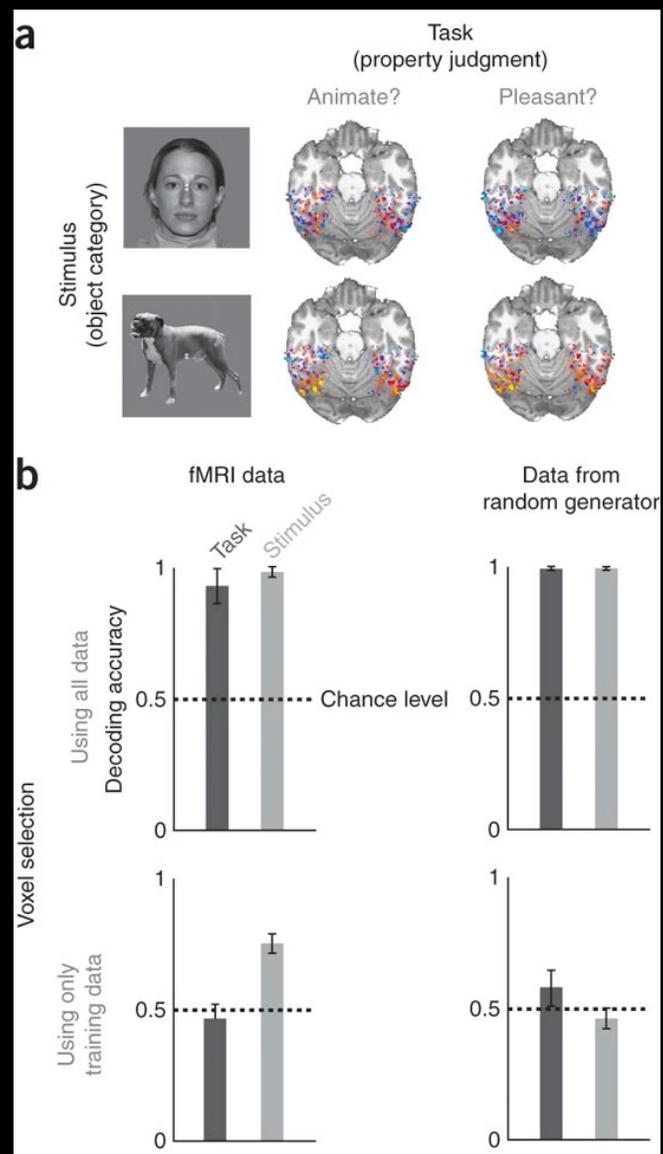
Demonstration

Kriegeskorte et al., 2010

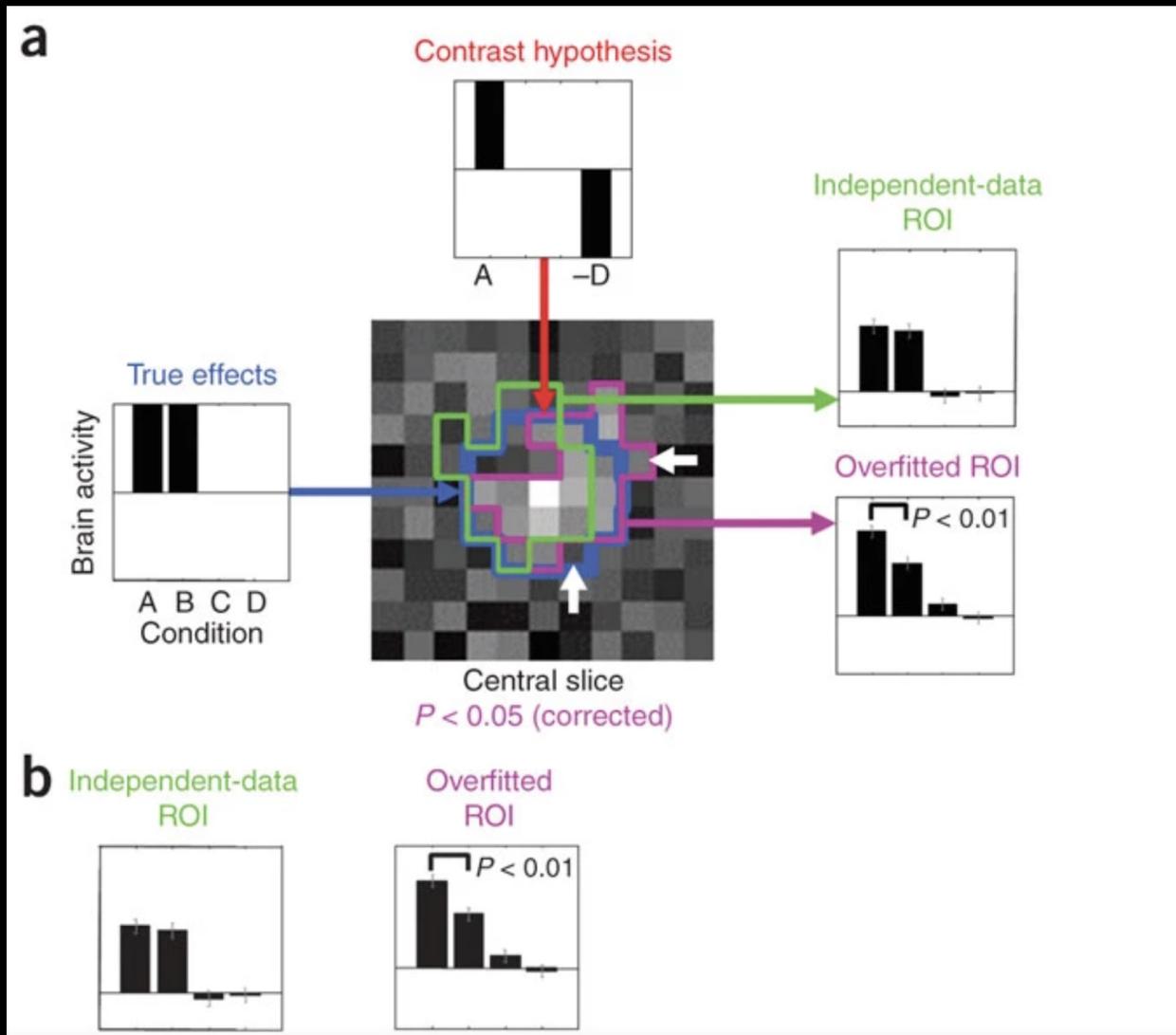
Recommendations for how to avoid non-independence



Kriegeskorte et al., 2010



Kriegeskorte et al., 2010



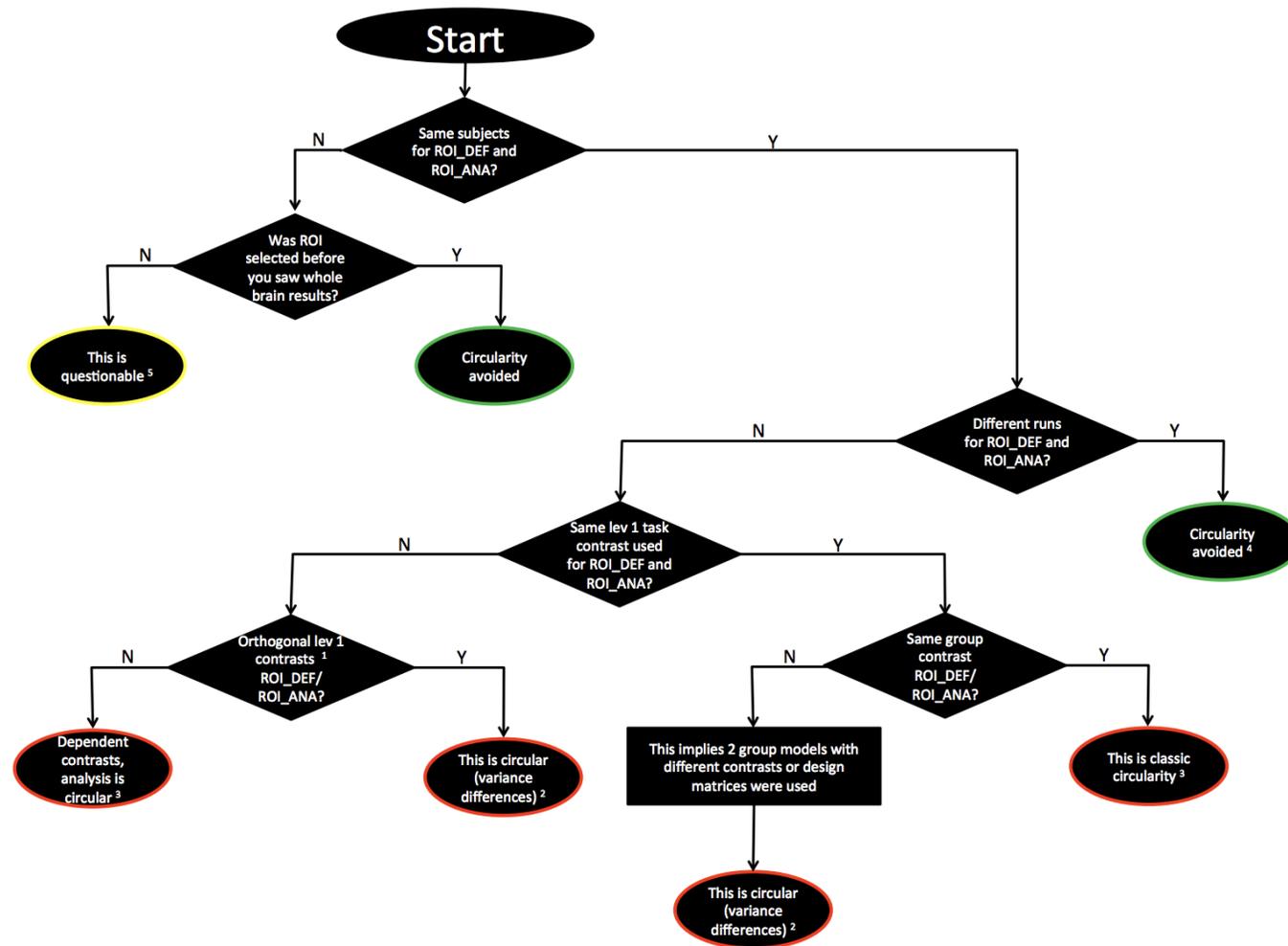
Kriegeskorte et al., 2010

The supplementary material reviews different scenarios

13. Can an omnibus F test safely be used to select channels for a subsequent selective analysis?

No. An omnibus F test determines whether the model as a whole explains significant variance in the data. It is, thus, sensitive to all effects modeled by the design matrix. Selection according to the omnibus F value will select channels whose data best conform to the model.

Consider the case of a single-predictor model (e.g. stimulation versus baseline). The omnibus F statistic will select channels that exhibit either positive or negative effects. Although there is no preference for either effect direction, channels with effect estimates close to zero will not be selected. If we were to test selected channels, t values under the null hypothesis would not follow a t distribution. In the extreme



ROI_DEF = ROI defining analysis; ROI_ANA = ROI analysis

- Orthogonal contrasts may refer to first level contrast vectors that are orthogonal (e.g. face+house and face-house) or orthogonal second level contrasts (group mean used for ROI_DEF where ROI_ANA is group comparison).
- See supplement of Kriegeskorte *et al.* (2009), bottom of p1 for orthogonal contrast and <http://mumfordbrainstats.tumblr.com/post/126904300281/less-obvious-double-dipping>. Typical practices are using a 1-sample t-test for ROI_DEF and a different ROI_ANA. For the example of using a group mean for ROI_DEF and 2-sample t for ROI_ANA, the contrasts are A-B and A+B, and $\text{Cov}(A-B, A+B) = \text{Var}(A) - \text{Var}(B)$, which is only 0 if the variances are equal. Thus orthogonality of the contrasts does not imply independence of the tests, unless the variances are equal. FYI, adding in a test of equal variances will not get you out of this jam, since it has its own false positive rate and those tests don't tend to work well.
- Kriegeskorte *et al.* (2009) or Vul *et al.* (2009).
- This assumes you either had 2 sets of runs with the same task or 2 sets of runs with different tasks, such as a localizer and task of interest. The latter is clearly independent and for the former, see page 2 of the supplement for Kriegeskorte *et al.* (2009) for a set of guidelines for splitting up runs for ROI_DEF and ROI_ANA.
- If you looked at the whole brain results that mirror what you'll do for ROI_ANA, even at uncorrected thresholds, it will now be difficult to choose an ROI from independent data without bias leaking in.

Lab Preview

Checking Data Quality

```
% Select all high resolution anatomical images
folder='C:\fMRI_Course\Data\SPM_Labs\Subjects\sub*\anatomy\precooked\wanat_hires.nii';
files=dir(folder);

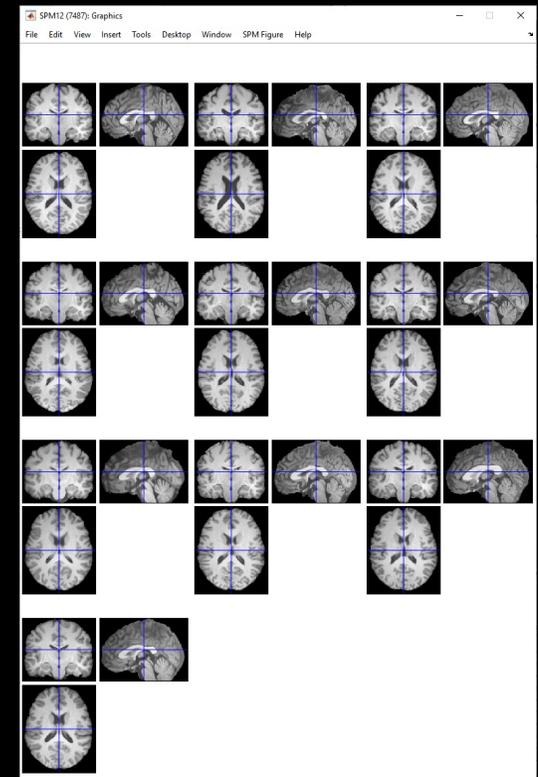
%Start with an empty variable in case we run this script multiple times
clear myHiRes

% Combine folder names with file names
for n=1:size(files);

    myHiRes{n} = [files(n).folder , '\', files(n).name , ',1' ];

end

% Transpose and convert to character array
myHiRes = char(myHiRes');
```



Lab Preview

Create Mean Anatomical

The image shows two windows from the SPM12 software. The left window, titled "SPM12 (7771): Menu", contains various processing options. The "ImCalc" button is highlighted with an orange box. An orange arrow points from this button to the right window. The right window, titled "Batch Editor", shows the "Image Calculator" module selected in the "Module List". The "Expression" field is highlighted with an orange box, and an orange arrow points from it to a smaller "Expression" dialog box. This dialog box contains the expression $(i1+i2+i3+i4+i5+i6)/6$. The "Batch Editor" also displays the "Current Module: Image Calculator" settings, including "Input Images" (6 files), "Output Filename", "Output Directory", and "Expression" (with a "<-X" button). The "Options" section includes "Data Matrix" (No - don't read images into data matrix), "Masking" (No implicit zero mask), "Interpolation" (Trilinear), and "Data Type" (INT16 - signed short). The "Expression" dialog box has "OK" and "Cancel" buttons. The "Batch Editor" also has a "Specify..." button at the bottom. At the bottom of the "Batch Editor" window, there is a section for "Expression" with example expressions: "* Mean of six images (select six images) (i1+i2+i3+i4+i5+i6)/6" and "* Make a binary mask image at threshold of 100 i1>100".

SPM12 (7771): Menu

Batch Editor

File Edit View SPM BasicIO

Module List

Image Calculator <-X

Current Module: Image Calculator

Help on: Image Calculator

Input Images 6 files

Output Filename

Output Directory

Expression <-X

Additional Variables

Options

. Data Matrix No - don't read images into data matrix

. Masking No implicit zero mask

. Interpolation Trilinear

. Data Type INT16 - signed short

Enter a value.

To clear a value, clear the input field and accept.

Leave input box with CTRL-TAB to access buttons.

$(i1+i2+i3+i4+i5+i6)/6$

OK Cancel

Specify...

Expression

Example expressions:

* Mean of six images (select six images)
 $(i1+i2+i3+i4+i5+i6)/6$

* Make a binary mask image at threshold of 100
 $i1>100$

SPM for functional MRI

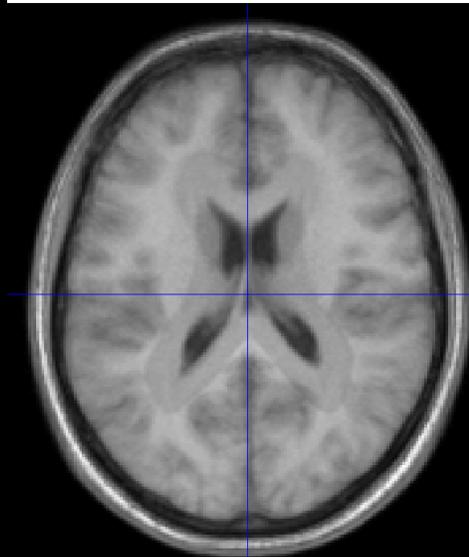
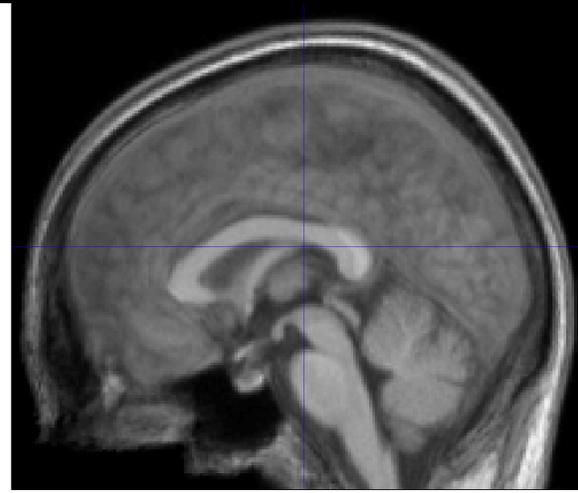
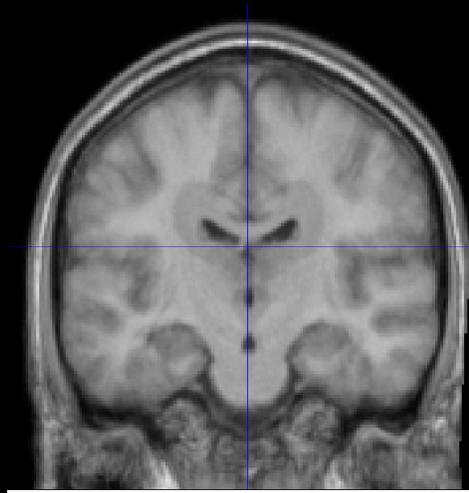
Display Check Reg Render... FMRI

Toolbox: PPIs ImCalc DICOM Import

Help Utils... Batch Quit

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Lab Preview



Lab Preview

Examine Results

